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FATTY ACIDS

(57) Abstract

By this invention, compositions and methods of use related to β -ketoacyl-ACP synthase of special interest are synthases obtainable from Cuphea species. Amino acid and nucleic acid for synthase protein factors are provided, as well as methods to utilize such sequences in constructs for production of genetically engineered plants having altered fatty acid compositions. Of particular interest is the expression of synthase protein factors in conjunction with expression of plant medium-chain acyl-ACP thioesterases for production of increased levels and/or modified ratios of medium-chain fatty acids in oils of transgenic plant seeds.

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PLANT FATTY ACID SYNTHASES AND USE IN IMPROVED METHODS FOR PRODUCTION OF MEDIUM-CHAIN FATTY ACIDS

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INTRODUCTION

Field of Invention

The present invention is directed to genes encoding plant fatty acid synthase enzymes relevant to fatty acid synthesis in plants, and to methods of using such genes in combination with genes encoding plant medium-chain preferring thioesterase proteins. Such uses provide a method to increase the levels of medium-chain fatty acids that may be produced in seed oils of transgenic plants.

Background

Higher plants synthesize fatty acids via a common metabolic pathway. In developing seeds, where fatty acids attached to triglycerides are stored as a source of energy for further germination, the fatty acid synthesis pathway is located in the plastids. The first step is the formation of acetyl-ACP (acyl carrier protein) from acetyl-CoA and ACP catalyzed by a short chain preferring condensing enzyme, ß-ketoacyl-ACP synthase (KAS) III. Elongation of acetyl-ACP to 16- and 18- carbon fatty acids involves the cyclical action of the following sequence of reactions: condensation with a two-carbon unit from malonyl-ACP to form a longer ß-ketoacyl-ACP (ß-ketoacyl-ACP synthase), reduction of the

keto-function to an alcohol (ß-ketoacyl-ACP reductase), dehydration to form an enoyl-ACP (ß-hydroxyacyl-ACP dehydrase), and finally reduction of the enoyl-ACP to form the elongated saturated acyl-ACP (enoyl-ACP reductase). ß-ketoacyl-ACP synthase I (KAS I), is primarily responsible for elongation up to palmitoyl-ACP (C16:0), whereas ß-ketoacyl-ACP synthase II (KAS II) is predominantly responsible for the final elongation to stearoyl-ACP (C18:0).

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Genes encoding peptide components of ß-ketoacyl-ACP synthases I and II have been cloned from a number of higher plant species, including castor (Ricinus communis) and Brassica species (USPN 5,510,255). KAS I activity was associated with a single synthase protein factor having an approximate molecular weight of 50 kD (synthase factor B) and KAS II activity was associated with a combination of two synthase protein factors, the 50 kD synthase factor B and a 46 kd protein designated synthase factor A. Cloning and sequence of a plant gene encoding a KAS III protein has been reported by Tai and Jaworski (Plant Physiol. (1993) 103:1361-1367).

The end products of plant fatty acid synthetase activities are usually 16- and 18-carbon fatty acids. There are, however, several plant families that store large

25 amounts of 8- to 14-carbon (medium-chain) fatty acids in their oilseeds. Recent studies with Umbellularia californica (California bay), a plant that produces seed oil rich in lauric acid (12:0), have demonstrated the existence of a medium-chain-specific isozyme of acyl-ACP thioesterase

in the seed plastids. Subsequent purification of the 12:0-ACP thioesterase from Umbellularia californica led to the cloning of a thioesterase cDNA which was expressed in seeds of Arabidopsis and Brassica resulting in a substantial accumulation of lauric acid in the triglyceride pools of these transgenic seeds (USPN 5,512,482). These results and subsequent studies with medium-chain thioesterases from other plant species have confirmed the chain-length-determining role of acyl-ACP thioesterases during de novo fatty acid biosynthesis (T. Voelker (1996) Genetic Engineering, Ed. J. K. Setlow, Vol. 18, pgs. 111-133).

DESCRIPTION OF THE FIGURES

Figure 1. DNA and translated amino acid sequence of Cuphea hookeriana KAS factor B clone chKAS B-2 are provided. Figure 2. DNA and translated amino acid sequence of Cuphea hookeriana KAS factor B clone chKAS B-31-7 are provided. Figure 3. DNA and translated amino acid sequence of Cuphea hookeriana KAS factor A clone chKAS A-2-7 are provided. Figure 4. DNA and translated amino acid sequence of Cuphea hookeriana KAS factor A clone chKAS A-1-6 are provided. Figure 5. DNA and translated amino acid sequence of Cuphea pullcherrima KAS factor B clone cpuKAS B/7-8 are provided. Figure 6. DNA and translated amino acid sequence of Cuphea pullcherrima KAS factor B clone cpuKAS B/8-7A are provided. 25 Figure 7. DNA and translated amino acid sequence of Cuphea pullcherrima KAS factor A clone cpuKAS A/p7-6A are provided. Figure 8. Preliminary DNA sequence of Cuphea pullcherrima KAS factor A clone cpuKAS A/p8-9A is provided.

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- Figure 9. DNA and translated amino acid sequence of Cuphea hookeriana KASIII clone chKASIII-27 are provided.
- Figure 10. The activity profile for purified cpuKAS B/8-7A using various acyl-ACP substrates is provided.
- Figure 11. The activity profile for purified chKAS A-2-7 and chKAS A-1-6 using various acyl-ACP substrates is provided.
 - Figure 12. The activity profile for purified castor KAS factor A using various acyl-ACP substrates is provided.
- Figure 13. The activity profile for purified castor KAS 10 factor B using various acyl-ACP substrates is provided. Figure 14. A graph showing the number of plants arranged according to C8:0 content for transgenic plants containing CpFatB1 versus transgenic plants containing CpFatB1 + chKAS A-2-7 is provided.
 - Figure 15. Graphs showing the %C10/%C8 ratios in transgenic plants containing ChFatB2 (4804-22-357) and in plants resulting from crosses between 4804-22-357 and 5401-9 (chKAS A-2-7 plants) are provided.
- Figure 16. Graphs showing the %C10 + %C8 contents in 20 transgenic plants containing ChFatB2 (4804-22-357) and in plants resulting from crosses between 4804-22-357 and 5401-9 (chKAS A-2-7 plants) are provided.
 - Figure 17. Graphs showing the %C10/%C8 ratios in transgenic plants containing ChFatB2 (4804-22-357) and in plants resulting from crosses between 4804-22-357 and 5413-17 (chKAS A-2-7 + CpFatB1 plants) are provided.
 - Figure 18. Graphs showing the %C10 + %C8 contents in transgenic plants containing ChFatB2 (4804-22-357) and in

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plants resulting from crosses between 4804-22-357 and 5413-17 (chKAS A-2-7 + CpFatB1 plants) are provided.

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Figure 19. Graphs showing the %C12:0 in transgenic plants containing Uc FatB1 (LA86DH186) and in plants resulting from crosses with wild type (X WT) and with lines expressing Ch KAS A-2-7.

Figure 20. Graph showing the relative proportions of C12:0 and C14:0 fatty acids in the seeds of transgenic plants containing Uc FatB1 (LA86DH186) and in plants resulting from crosses with wild type (X WT) and with lines expressing Ch KAS A-2-7.

Figure 21. Graphs showing the %C18:0 in transgenic plants containing Garm FatB1 (5266) and in seeds of plants resulting from crosses with wild type (X WT) and with lines expressing Ch KAS A-2-7.

Figure 22. The activity profile of Ch KAS A in protein extracts from transgenic plants containing Ch KAS A-2-7. Extracts were preptreated with the indicated concentrations of cerulenin.

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SUMMARY OF THE INVENTION

By this invention, compositions and methods of use related to ß-ketoacyl-ACP synthase (KAS) are provided. Also of interest are methods and compositions of amino acid and nucleic acid sequences related to biologically active plant synthase(s).

In particular, genes encoding KAS protein factors A and B from Cuphea species are provided. The KAS genes are of interest for use in a variety of applications, and may be

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used to provide synthase I and/or synthase II activities in transformed host cells, including bacterial cells, such as E. coli, and plant cells. Synthase activities are distinguished by the preferential activity towards longer and shorter acyl-ACPs as well as by the sensitivity towards the KAS specific inhibitor, cerulenin. Synthase protein preparations having preferential activity towards medium chain length acyl-ACPs are synthase I-type or KAS I. The KAS I class is sensitive to inhibition by cerulenin at concentrations as low as 1 µM. Synthases having preferential activity towards longer chain length acyl-ACPs are synthase II-type or KAS II. The KAS enzymes of the II-type are also sensitive to cerulenin, but at higher concentrations (50µM). Synthase III-type enzymes have preferential activity towards short chain length acyl-ACPs and are insensitive to cerulenin inhibition.

Nucleic acid sequences encoding a synthase protein may be employed in nucleic acid constructs to modulate the amount of synthase activity present in the host cell,

20 especially the relative amounts of synthase I-type, synthase II-type and synthase III-type activity when the host cell is a plant host cell. In addition, nucleic acid constructs may be designed to decrease expression of endogenous synthase in a plant cell as well. One example is the use of an antisense synthase sequence under the control of a promoter capable of expression in at least those plant cells which normally produce the enzyme.

Of particular interest in the present invention is the coordinate expression of a synthase protein with the

expression of thioesterase proteins. For example, coordinated expression of synthase factor A and a medium-chain thioesterase provides a method for increasing the level of medium-chain fatty acids which may be harvested from transgenic plant seeds. Furthermore, coordinated expression of a synthase factor A gene with plant medium-chain thioesterase proteins also provides a method by which the ratios of various medium-chain fatty acids produced in a transgenic plant may be modified. For example, by expression of a synthase factor A, it is possible to increase the ratio of C10/C8 fatty acids which are produced in plant seed oils as the result of expression of a thioesterase having activity on C8 and C10 fatty acids.

DETAILED DESCRIPTION OF THE INVENTION

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A plant synthase factor protein of this invention includes a sequence of amino acids or polypeptide which is required for catalyzation of a condensation reaction between an acyl-ACP having a chain length of C2 to C16 and malonyl-ACP in a plant host cell. A particular plant synthase factor protein may be capable of catalyzing a synthase reaction in a plant host cell (for example as a monomer or homodimer) or may be one component of a multiple peptide enzyme which is capable of catalyzing a synthase reaction in a plant host cell, i.e. one peptide of a heterodimer.

Synthase I (KAS I) demonstrates preferential activity towards acyl-ACPs having shorter carbon chains, C_2 - C_{14} and is sensitive to inhibition by cerulenin at concentrations of 1 μ M. Synthase II (KAS II) demonstrates preferential

activity towards acyl-ACPs having longer carbon chains, C_{14} - C_{16} , and is inhibited by concentrations of cerulenin (50 μ M). Synthase III demonstrates preferential activity towards acyl-CoAs having very short carbon chains, C_{2} to C_{6} , and is insensitive to inhibition by cerulenin.

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Synthase factors A and B, and synthase III proteins obtained from medium-chain fatty acid producing plant species of the genus Cuphea are described herein. As described in the following Examples, synthase A from C. hookeriana is naturally expressed at a high level and only in the seeds. C. hookeriana synthase B is expressed at low levels in all tissues examined. Expression of synthase A and synthase B factors in E. coli and purification of the resulting proteins is employed to determine activity of the various synthase factors. Results of these analyses indicate that synthase factor A from Cuphea hookeriana has the greatest activity on 6:0-ACP substrates, whereas synthase factor B from Cuphea pullcherrima has greatest activity on 14:0-ACP. Similar studies with synthase factors A and B from castor demonstrate similar activity profiles between the factor B synthase proteins from Cuphea and The synthase A clone from castor, however, castor. demonstrates a preference for 14:0-ACP substrate.

Expression of a Cuphea hookeriana KAS A protein in

25 transgenic plant seeds which normally do not produce mediumchain fatty acids does not result in any detectable
modification of the fatty acid types and contents produced
in such seeds. However, when Cuphea hookeriana KAS A
protein is expressed in conjunction with expression of a

medium-chain acyl-ACP thioesterase capable of providing for production of C8 and C10 fatty acids in plant seed oils, increases in the levels of medium-chain fatty acids over the levels obtainable by expression of the medium-chain thioesterase alone are observed. In addition, where significant amounts of C8 and C10 fatty acids are produced as the result of medium-chain thioesterase expression, co-expression of a Cuphea KAS A protein also results in an alteration of the proportion of the C8 and C10 fatty acids that are obtained. For example, an increased proportion of C10 fatty acids may be obtained by co-expression of Cuphea hookeriana ChFatB2 thioesterase and a chKAS A synthase factor proteins.

Furthermore, when Cuphea hookeriana KAS A protein is expressed in conjunction with expression of a medium-chain acyl-ACP thioesterase capable of providing for production of C12 fatty acids in plant seed oils, increases in the levels of medium-chain fatty acids over the levels obtainable by expression of the medium-chain thioesterase alone are also observed. In addition, where significant amounts of C12 and C14 fatty acids are produced as the result of medium-chain thioesterase expression, co-expression of a Cuphea KAS A protein also results in an alteration of the proportion of the C12 and C14 fatty acids that are obtained. For example, an increased proportion of C12 fatty acids may be obtained by co-expression of Uc FatB1 thioesterase and a chKAS A synthase factor proteins.

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However, when Cuphea hookeriana KAS A protein is expressed in conjunction with the expression of a long-chain

acyl-ACP thioesterase capable of providing for production of C18 and C18:1 fatty acids in plant seed oils, no effect on the production of long chain fatty acids was observed.

Furthermore, when plants transformed to express a long chain acyl-ACP thioesterase from mangosteen (GarmFatAl, U.S. Patent Application No. 08/440,845), which preferentially hydrolyzes C18:0 and C18:1 fatty acyl-ACPs, are crossed with nontransformed control plants, a significant reduction in the levels of C18:0 is obtained. Similar reductions are also observed in the levels of C18:0 in the seeds of plants resulting from crosses between plants transformed to express the GarmFatAl and plants expressing the Cuphea hookeriana KAS A protein.

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Thus, the instant invention provides methods of increasing and/or altering the medium-chain fatty acid 15 compositions in transgenic plant seed oils by co-expression of medium-chain acyl-ACP thioesterases with synthase factor proteins. Furthermore, various combinations of synthase factors and medium-chain thioesterases may be achieved depending upon the particular fatty acids desired. For 20 example, for increased production of C14 fatty acids, synthase protein factors may be expressed in combination with a C14 thioesterase, for example from Cuphea palustris or nutmeg may be employed (WO 96/23892). In addition, thioesterase expression may be combined with a number of 25 different synthase factor proteins for additional effects on medium-chain fatty acid composition.

Synthases of use in the present invention include modified amino acid sequences, such as sequences which have

been mutated, truncated, increased and the like, as well as such sequences which are partially or wholly artificially synthesized. The synthase protein encoding sequences provided herein may be employed in probes for further screening or used in genetic engineering constructs for transcription or transcription and translation in host cells, especially plant host cells. One skilled in the art will readily recognize that antibody preparations, nucleic acid probes (DNA and RNA) and the like may be prepared and used to screen and recover synthases and/or synthase nucleic acid sequences from other sources. Typically, a homologously related nucleic acid sequence will show at least about 60% homology, and more preferably at least about 70% homology, between the R. communis synthase and the given plant synthase of interest, excluding any deletions which may be present. Homology is determined upon comparison of sequence information, nucleic acid or amino acid, or through hybridization reactions.

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Recombinant constructs containing a nucleic acid

sequence encoding a synthase protein factor or nucleic acid

sequences encoding a synthase protein factor and a mediumchain acyl-ACP thioesterase may be prepared by methods well
known in the art. Constructs may be designed to produce
synthase in either prokaryotic or eukaryotic cells. The

increased expression of a synthase in a plant cell,
particularly in conjunction with expression of medium-chain
thioesterases, or decreasing the amount of endogenous
synthase observed in plant cells are of special interest.

Synthase protein factors may be used, alone or in combination, to catalyze the elongating condensation reactions of fatty acid synthesis depending upon the desired result. For example, rate influencing synthase activity may reside in synthase I-type, synthase II-type, synthase IIItype or in a combination of these enzymes. Furthermore, synthase activities may rely on a combination of the various synthase factors described herein.

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Constructs which contain elements to provide the transcription and translation of a nucleic acid sequence of 10 interest in a host cell are "expression cassettes". Depending upon the host, the regulatory regions will vary, including regions from structural genes from viruses, plasmid or chromosomal genes, or the like. For expression in prokaryotic or eukaryotic microorganisms, particularly unicellular hosts, a wide variety of constitutive or regulatable promoters may be employed. Among transcriptional initiation regions which have been described are regions from bacterial and yeast hosts, such as E. coli, B. subtilis, Saccharomyces cerevisiae, including genes such 20 as ß-galactosidase, T7 polymerase, trp-lac (tac), trp E and the like.

An expression cassette for expression of synthase in a plant cell will include, in the 5' to 3' direction of transcription, a transcription and translation initiation 25 control regulatory region (also known as a "promoter") functional in a plant cell, a nucleic acid sequence encoding a synthase, and a transcription termination region. Numerous transcription initiation regions are available

which provide for a wide variety of constitutive or regulatable, e.g., inducible, transcription of the desaturase structural gene. Among transcriptional initiation regions used for plants are such regions associated with cauliflower mosaic viruses (35S, 19S), and structural genes such as for nopaline synthase or mannopine synthase or napin and ACP promoters, etc. The transcription/ translation initiation regions corresponding to such structural genes are found immediately 5' upstream to the respective start codons. Thus, depending upon the intended use, different promoters may be desired.

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Of special interest in this invention are the use of promoters which are capable of preferentially expressing the synthase in seed tissue, in particular, at early stages of seed oil formation. Examples of such seed-specific promoters include the region immediately 5' upstream of a napin or seed ACP genes such as described in USPN 5,420,034, desaturase genes such as described in Thompson et al (Proc. Nat. Acad. Sci. (1991) 88:2578-2582), or a Bce-4 gene such as described in USPN 5,530,194. Alternatively, the use of the 5' regulatory region associated with the plant synthase structural gene, i.e., the region immediately 5' upstream to a plant synthase structural gene and/or the transcription termination regions found immediately 3' downstream to the plant synthase structural gene, may often be desired. general, promoters will be selected based upon their expression profile which may change given the particular application.

In addition, one may choose to provide for the transcription or transcription and translation of one or more other sequences of interest in concert with the expression or anti-sense of the synthase sequence, particularly medium-chain plant thioesterases such as described in USPN 5,512,482, to affect alterations in the amounts and/or composition of plant oils.

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When one wishes to provide a plant transformed for the combined effect of more than one nucleic acid sequence of interest, a separate nucleic acid construct may be provided for each or the constructs may both be present on the same plant transformation construct. The constructs may be introduced into the host cells by the same or different methods, including the introduction of such a trait by crossing transgenic plants via traditional plant breeding methods, so long as the resulting product is a plant having both characteristics integrated into its genome.

Normally, included with the DNA construct will be a structural gene having the necessary regulatory regions for expression in a host and providing for selection of transformed cells. The gene may provide for resistance to a cytotoxic agent, e.g. antibiotic, heavy metal, toxin, etc., complementation providing prototrophy to an auxotrophic host, viral immunity or the like. Depending upon the number of different host species into which the expression construct or components thereof are introduced, one or more markers may be employed, where different conditions for selection are used for the different hosts.

The manner in which the DNA construct is introduced into the plant host is not critical to this invention. Any method which provides for efficient transformation may be employed. Various methods for plant cell transformation include the use of Ti- or Ri-plasmids, microinjection, electroporation, liposome fusion, DNA bombardment or the like. In many instances, it will be desirable to have the construct bordered on one or both sides by T-DNA, particularly having the left and right borders, more particularly the right border. This is particularly useful when the construct uses A. tumefaciens or A. rhizogenes as a mode for transformation, although the T-DNA borders may find use with other modes of transformation.

The expression constructs may be employed with a wide

15 variety of plant life, particularly plant life involved in

the production of vegetable oils. These plants include, but

are not limited to rapeseed, peanut, sunflower, safflower,

cotton, soybean, corn and oilseed palm.

explants may be combined and incubated with the transformed Agrobacterium for sufficient time for transformation, the bacteria killed, and the plant cells cultured in an appropriate selective medium. Once callus forms, shoot formation can be encouraged by employing the appropriate plant hormones in accordance with known methods and the shoots transferred to rooting medium for regeneration of plants. The plants may then be grown to seed and the seed used to establish repetitive generations and for isolation of vegetable oils.

The invention now being generally described, it will be more readily understood by reference to the following examples which are included for purposes of illustration only and are not intended to limit the present invention.

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EXAMPLES

Example 1 Cuphea KAS Factor A and B Gene Cloning

Total RNA isolated from developing seeds of Cuphea hookeriana and Cuphea pullcherrima was used for cDNA synthesis in commercial 1-based cloning vectors. cloning each type of KAS gene, approximately 400,000-500,000 unamplified recombinant phage were plated and the plagues transferred to nitrocellulose. For KAS factor B cloning from C. hookeriana, a mixed probe containing Brassica napus KAS factor B and Ricinus communis (Castor) KAS factor B radiolabeled cDNA's was used. Similarly, a mixed probe containing Brassica napus KAS factor A and Ricinus communis KAS factor A cDNA clones was used to obtain C. hookeriana KAS factor A genes. For KASIII, a spinach KASIII cDNA clone obtained from Dr. Jan Jaworski was radiolabeled and used as a probe to isolate a KASIII clone from C. hookeriana. For KAS B and KAS A cloning from C. pullcherrima, C. hookeriana KAS B and KAS A genes chKAS B-2 and chKAS A-2-7 (see below) were radiolabeled and used as probes.

DNA sequence and translated amino acid sequence for Cuphea KAS clones are provided in Figures 1-9. Cuphea hookeriana KAS factor B clones chKAS B-2 and chKAS B-31-7

are provided in Figures 1 and 2. Neither of the clones is full length. Cuphea hookeriana KAS Factor A clones chKAS A-2-7 and chKAS A-1-6 are provided in Figures 3 and 4. chKAS A-2-7 contains the entire encoding sequence for the KAS factor protein. Based on comparison with other plant synthase proteins, the transit peptide is believed to be represented in the amino acids encoded by nucleotides 125-466. chKAS A-1-6 is not a full length clone although some transit peptide encoding sequence is present. Nucleotides 1-180 represent transit peptide encoding sequence, and the mature protein encoding sequence is believed to begin at nucleotide 181.

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Cuphea pullcherrima KAS factor B clones cpuKAS B/7-8 and cpuKAS B/8-7A are provided in Figures 5 and 6. Both of the clones contain the entire encoding sequences for the KAS 15 factor B proteins. The first 35 amino acids of cpuKAS B/7-8 are believed to represent the transit peptide, with the mature protein encoding sequence beginning at nucleotide The first 39 amino acids of cpuKAS B/8-7A are believed 233. to represent the transit peptide, with the mature protein 20 encoding sequence beginning at nucleotide 209. Cuphea pullcherrima KAS factor A clones cpuKAS A/p7-6A and cpuKAS A-p8-9A are provided in Figures 7 and 8. Both of the clones contain the entire encoding sequences for the KAS factor A proteins. Translated amino acid sequence of cpuKAS A/p7-6A 25 is provided. The mature protein is believed to begin at the lysine residue encoded 595-597, and the first 126 amino acids are believed to represent the transit peptide. The DNA sequence of KAS A clone cpuKAS A-p8-9A is preliminary.

Further analysis will be conducted to determine final DNA sequence and reveal the amino acid sequence encoded by this gene.

DNA and translated amino acid sequence of Cuphea hookeriana KASIII clone chKASIII-27 is provided in Figure 9. The encoding sequence from nucleotides 37-144 of chKASIII-27 are believed to encode a transit peptide, and the presumed mature protein encoding sequence is from nucleotides 145-1233.

10 Deduced amino acid sequence of the C. hookeriana KAS factor B and KAS factor A cDNA's reveals strong homology to the Brassica napus and Ricinus communis clones previously reported. The C. hookeriana KAS factor B clone is more homologous to the Ricinus and Brassica KAS factor B clones (94% and 91% respectively) than it is to the Ricinus and 15 Brassica KAS factor A clones (60% for both). Furthermore, the C. hookeriana KAS factor A clone is more homologous to the Ricinus and Brassica KAS factor A clones (85% and 82% respectively) than it is the Ricinus and Brassica KAS factor B clone (60% for both). The C. hookeriana KAS factor B 20 cDNAs designated as chKAS B-2 and chKAS B-31-7 are 96% identical within the mature portion of the polypeptide. Similarly, the deduced amino acid sequence of the mature protein regions of the C. hookeriana KAS factor A clones chKAS A-2-7 and chKAS A-1-6 are 96% identical. The C. 25 pullcherrima KAS clones also demonstrate homology to the R. communis and Brassica napus KAS clones. The mature protein portion of all of the KAS factor A family members in the different Cuphea species are 95% identical. Similarly the

mature protein portion of the KAS factor B genes in Cuphea are also 95-97% identical with each other. However there is only approximately 60% sequence identity between KAS factor B and KAS factor A clones either within the same or different species of Cuphea.

Example 2 Levels and Patterns of Expression

mRNA and its abundance on the blot.

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To examine tissue specificity of KAS expression in Cuphea hookeriana, Northern blot analysis was conducted using total RNA isolated from seed, root, leaf and flower tissue. Two separate but identical blots were hybridized with either chKAS B-31-7 or chKAS A-2-7 coding region probes. The data from this RNA blot analysis indicate that KAS B is expressed at a similar level in all tissues examined, whereas KAS A expression is detected only in the These results also demonstrate a different level of expression for each of the synthases. KAS A is an abundant message, whereas KAS B is expressed at low levels. Furthermore, even under highly stringent hybridization conditions (65_C, 0.1 X SSC, 0.5% SDS), the KAS A probe hybridizes equally well with two seed transcripts of 2.3 and The larger hybridizing band is likely the 1.9 kb. transcript of the KAS A-2-7 gene since the size of its cDNA is 2046bp, and the number of clones obtained from cDNA screening corresponds well with the apparent mobility of the

Example 3 Expression of Plant KAS Genes in E.coli

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DNA fragments encoding the mature polypeptide of the Cuphea hookeriana KAS A cDNAs and the Cuphea pullcherrima KAS B cDNAs were obtained by PCR and cloned into a QIAexpress expression vector (Qiagene). Experimental conditions for maximum level of expression were determined for all of these clones and the parameters for highest level of soluble fraction were identified. Cells are grown in ECLB media containing 1M sorbitol and 2.5 mM betaine overnight and subcultured as a 1:4 dilution in the same medium. Cells are then grown for 2 hours (to approximately .6-.8 O.D.) and induced with 0.4 mM IPTG and allowed to grow for 5 more hours.

Enzyme activity of the affinity purified recombinant enzymes obtained from over-expression of the chKAS A-2-7 and cpuKAS B/8-7A clones was measured using a wide range of acyl-ACP substrates (6:0- to 16:1-ACP). The activity profile for cpuKAS B/8-7A is provided in Fig.10. The data demonstrate that the enzyme is active with all acyl-ACP substrates examined, although activity on 6:0 to 14:0-ACP substrates is substantially greater than the activity on 16:0 and 16:1 substrates.

The activity profile of the *C. hookeriana* KAS A clones

25 chKAS A-2-7 and chKAS A-1-6 is provided in Figure 11. The *C. hookeriana* KAS A clones are most active with C:6, and have the least activity with C:16:0 substrates. However, the activity of this clone on even the preferred C6:0 substrate

is 50 fold lower than the activity of the *C. pullcherrima* KAS B clones.

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A fragment containing the mature protein encoding portion of a R. communis KAS factor A clone was also cloned into a QIAexpress expression vector, expressed in E. coli and the enzyme affinity purified as described above. The activity profile for castor KAS A is provided in Figure 12. Highest activity is observed with C14:0 substrates, although some activity is also seen with C6:0 and C16:1. In comparison, the activity profile obtained from purified R. communis KAS factor B also using the QIAexpress expression system is provided in Figure 13. The KAS B clone demonstrates substantially higher levels of activity (10 fold and higher) than the R. communis KAS A clone. The preference of the KAS factor B for 6:0- to 14:0-ACP substrates is consistent with the previous observations that this protein provides KAS I activity.

Example 4 KAS and TE Expression in Transgenic Seed

Dehesh et al. (1996) Plant Physiol. 110:203-210) and the chKAS A-2-7 were PCR amplified, sequenced, and cloned into a napin expression cassette. The napin/cp FatB1 and the napin/KAS A-2-7 fusions were ligated separately into the binary vector pCGN1558 (McBride and Summerfelt (Pl.Mol.Biol. (1990) 14:269-276) and transformed into A. tumefaciens, EHA101. The resulting CpFatB1 binary construct is pCGN5400 and the chKAS A-2-7 construct is pCGN5401. Agrobacterium mediated transformation of a Brassica napus canola variety

was carried out as described by Radke et al. (Theor. Appl. Genet. (1988) 75:685-694; Plant Cell Reports (1992) 11:499-505). Several transgenic events were produced for each of the pCGN5400 and pCGN5401 constructs.

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A double gene construct containing a napin/cpFatB1 expression construct in combination with a napin/chKAS A-2-7 expression construct was also assembled, ligated into a binary vector and used for co-cultivation of a canola Brassica variety. The binary construct containing the chFatB1 and chKAS A-2-7 expression constructs is pCGN5413.

Fatty acid analysis of 26 transgenic lines containing chKAS A-2-7 (5401 lines) showed no significant changes in the oil content or profile as compared to similar analyses of wild type canola seeds of the transformed variety.

Fatty acid analysis of 36 transgenic lines containing cpFatB1 (5400 lines) showed increased levels of C:8 and C:10 in transgenic seeds. The highest level of C:8 observed in a pool seed sample was 4.2 mol%. The C:10 levels were between 30 and 35% of the C:8 content. Fatty acid analysis of 25 transgenic lines containing the TE/KAS A tandem (5413 lines) demonstrated an overall increase in both C:8 and C:10 levels relative to those observed with TE containing lines (5400) alone. In lines containing the cpFatB1 construct alone, the average level of C:8 average were 1.5 mol%, whereas the C:8 average levels in TE/KAS A tandem containing lines was 2.37 mol%. The ratio of C:8 to C:10 remained constant in both populations. The number of transgenic events relative to the C:8 content are presented in Figure 14. These data show that the transgenic events with tandem TE/KAS A construct

yield more lines with higher levels of C:8 than those events with single TE construct. For example, several lines containing nearly 7 mole% C8 were obtained with the TE/KAS A pCGN5413 construct, whereas the highest C8 containing line from the pCGN5400 TE alone transformation contained 4.2 mole% C8.

Half seed analysis of the T3 generation of transgenic canola plants expressing a ChFatB2 (*C. hookeriana* thioesterase; Dehesh et al. (1996) The Plant Journal 9:167-172) indicate that these plant can accumulate up to 22 weight% (33 mol%) of 8:0 and 10:0 fatty acids (4804-22-357). Segregation analysis shows that these transformants contain two loci and that they are now homozygous. Selected plants grown from these half seeds were transferred into the greenhouse and later crossed with T1 transformants that had been transformed with either Cuphea hookeriana KAS A (5401) alone or KAS A/CpFatB1 double constructs (5413).

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Fatty acid analysis of several events resulting from the crosses between transgenic lines containing ChFatB2

20 (4804-22-357) and chKAS A-2-7 (5401-9), reveal an increase in the ratio of C:10/C:8 levels (Figure 15). This C:10/C:8 ratio in nearly all of the transgenic events containing ChFatB2 TE alone fluctuates between 3 and 6, whereas in the F1 generation of transgenic containing both the TE and the C:10 levels is accompanied by an increase in the total C:8 and C:10 content (Figure 16). The sum of the C:8 and C:10 fatty acids in the heterozygous F1 lines is as high as those in the homozygous parent line (4804-22-357), whereas the

heterozygous lines usually contain substantially less C:8 and C:10 than the homozygous lines.

Similar results were observed in F1 generation seeds resulting from crosses performed between 4804-22-357 (ChFatB2) and the 5413-17 event (CpFatB1 and chKAS A-2-7 5 tandem). Levels of C:8 and C:10 in the 5413-17 line were 6.3 and 2.8 mol% respectively. Data presented in Figure 17 show that there is shift towards C:10 fatty acids as was observed with the 4804-22-357 (ChFatB2) x 5401-9 (chKAS A-2-7) crosses. Furthermore, Figure 18 indicates the presence 10 of two separate populations of heterozygotes. Those containing approximately 9-11 weight percent C:10 + C:8 are believed to represent offspring containing a single copy of the ChFatB1 TE gene and no copies of the CpFatB1 and chKAS A genes from 5413. Those plants containing approximately 15-15 20 weight percent C:10 + C:8 are believed to represent the heterozygotes containing a single ChFatB1 TE gene as well as the CpFatB1 and chKAS A genes from 5413. Thus, the level of the C:10 + C:8 fatty acids does not decrease to 50% of that detected in parent lines when a copy of the ChKAS A gene is 20 present.

To further characterize the chain length specificity of the Cuphea hookeriana KAS A enzyme, crosses between transgenic Brassica napus lines containing a California Bay (Umbellularia californica) 12:0 specific thioesterase, Uc FatB1 (USPN 5,344,771) and chKAS A-2-7 (5401-9) were made. Half seed analysis of transgenic plants containing Uc fatB1 have previuosly indicated that these plants can accumulate up to 52 mol% C12:0 in the seed oil of homozygous dihaploid

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lines (LA86DH186). Crosses between the line LA86DH186 and untransformed control *Brassica* demonstrated a decrease in the C12:0 levels.

However, crosses between LA86DH186 and the 5401-9 hemizygous line led to an accumulation of up to 57 mol% C12:0 in the seed oil of F1 progeny (Figure 19). Interestingly, in crosses with LA86DH186 x untransformed control line and LA86DH186 x 5401-9, levels of C14:0 in the seeds of the F1 progeny decreased to 50% of the levels obtained in homozygous LA86DH186 lines (Figure 20). 10 Furthermore, increases in the proportion of C12:0 fatty acid resulted in a substantial decline in the proportions of all the long-chain fatty acyl groups (C16:0, C18:0, C18:2, and C18:3). These results indicate that the ChKAS A-2-7 is an enzyme with substrate specificity ranging from C6:0 to 15 C10:0-ACP, and that its over-expression ultimately reduces the longer chain acyl-ACP pools.

Further evidence is obtained in support of the chain length specificity of the ChKAS A-2-7 in crosses of the 5401-9 line with a transgenic line (5266) expressing an 18:1/18:0 TE from Garcinia mangostana (GarmFatA1, US patent application No. 08/440,845). Transgenic Brassica line 5266 has been shown to accumulate up to 24 mol% Cl8:0 in the seed oil of homozygous lines (Figure 21). However, in the seed oil of F1 progeny of crosses between 5266 and 5401-9 levels of Cl8:0 were reduced to approximately 12 mol%. Furthermore, levels of Cl6:0 generated from these crosses was similar to the levels obtained from the seed oil of nontransgenic control plants.

Example 5 In vitro Analysis of Plant KAS Enzymes

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Seed extracts were prepared from developing seeds of nontransgenic controls or transgenic Brassica expressing chKAS A-2-7 as described in Slabaugh et al. (Plant Journal, 1998 in press) and Leonard et al. (Plant Journal, 1998, in In vitro fatty acid synthesis assays were performed press). as described by Post-Beittenmiller (J. Biol. Chem. (1991), 266:1858-1865). Extracts were concentrated by ammonium sulfate precipitation and desalting using P-6 columns (Bio-Rad, Hercules, CA). Reactions (65µl) contained 0.1M 10 Tris/HCl (pH 8.0), 1 mM dithiothreitol, 25 mM recombinant spinach ACP1, 1 mM NADH, 2 mM NADPH, 50 µM malonyl-CoA, 10 μ M [1-14C]acetyl-CoA (50 mCi/mmol), lmg/ml BSA, and 0.25 mg/ml seed protein. Selected seed extracts were preincubated with cerulenin at 23°C for 10 min. Reaction products were separated on an 18% acrlamide gel containing 2.25M urea, electroblotted onto to nitrocellulose and quntitated by phosporimaging using Image QuaNT software (Molecular Dynamics, Sunnyvale, CA). Authentic acyl-ACPs were run in parallel, immunoblotted and finally detected by anti-ACP serum to confirm fatty acid chain lengths.

The results (Figure 22) indicate that the fatty acid synthesis capabilities of transgenic Brasica (5401-9) seed extracts was greater than that obtained from in the nontransgenic controls as measured by the relative abundance 25 of C8:0- and C10:0-ACP at all time points tested. addition, pretreatment of the extracts with cerulenin, markedly reduced the synthesis of longer chain fatty acids in both the transgenic and nontransgenic control seed

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extracts. However, the extension of the spinach-ACP was much less inhibited in the seed extracts from the transgenic lines than in the seed extracts of nontransgenic control Brassica.

These data further support that Ch KAS A-2-7 is a condensing enzyme active on medium chain acyl-ACPs, and that expression of this enzyme in plants results in enlarged substrate pools to be hydrolyzed by medium-chain specific thioesterases. Furthermore, these data suggest that chKAS A-2-7 also is a cerulenin-resistant condensing enzyme.

All publications and patent applications mentioned in this specification are indicative of the level of skill of those skilled in the art to which this invention pertains.

- All publications and patent applications are herein incorporated by reference to the same extent as if each individual publication or patent application was specifically and individually indicated to be incorporated by reference.
- Although the foregoing invention has been described in some detail by way of illustration and example for purposes of clarity of understanding, it will be obvious that certain changes and modifications may be practiced within the scope of the appended claim.

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MISSING UPON TIME OF PUBLICATION

- 13. The construct of Claim 5 wherein said encoding sequence is cpuKAS A/p8-9A.
- 14. The construct of Claim 5 wherein said encoding sequence is chKASIII-27.
- 15. An improved method for producing medium-chain fatty acids in transgenic plant seeds by expression of a plant medium-chain thioesterase protein heterologous to said transgenic plant,

the improvement comprising expression of a plant synthase

factor protein heterologous to said transgenic plant in

conjunction with expression of said plant medium-chain

thioesterase, whereby the percentage of medium-chain fatty

acids produced in seeds expressing both a plant synthase factor

protein and a plant medium-chain thioesterase protein is

increased as compared to the percentage of medium-chain fatty

acids produced in seeds expressing only said plant medium-chain

thioesterase protein.

- 16. The method of Claim 15 wherein said medium-chain thioesterase protein is a ChFatB2 protein.
- 17. The method of Claim 15 wherein said medium-chain thioesterase protein is a CpFatB1 protein.
 - 18. The method of Claim 15 wherein said medium-chain thioesterase protein is a C12 preferring thioesterase from California bay.
- 19. The method of Claim 15 wherein said plant synthase factor protein is expressed from a construct according to Claim 1.
 - 20. The method of Claim 19 wherein said synthase factor A protein is from a Cuphea species.

21. The method of Claim 20 wherein said Cuphea species is C. hookeriana or C. pullcherrima.

22. A method of altering the medium-chain fatty acid composition in plant seeds expressing a heterologous plant medium-chain preferring thioesterase, wherein said method comprises

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providing for expression of a plant synthase factor protein heterologous to said transgenic plant in conjunction with expression of a plant medium-chain thioesterase protein heterologous to said transgenic plant, whereby the composition of medium-chain fatty acids produced in said seeds is modified as compared to the composition of medium-chain fatty acids produced in seeds expressing said plant medium-chain thioesterase protein in the absence of expression of said plant synthase factor protein.

- 23. The method of Claim 22 wherein said medium-chain thioesterase protein is a ChFatB2 protein.
- 24. The method of Claim 22 wherein said medium-chain thioesterase protein is a CpFatB1 protein.
- 25. The method of Claim 22 wherein said medium-chain thioesterase protein is a C12 preferring thioesterase from California bay.
 - 26. The method of Claim 22 wherein said plant synthase factor protein is expressed from a construct according to Claim 1.
 - 27. The method of Claim 26 wherein said synthase factor A protein is from a Cuphea species.
 - 28. The method of Claim 27 wherein said Cuphea species is C. hookeriana or C. pullcherrima.

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29. The method of Claim 22 wherein said fatty acid composition is enriched for C10 fatty acids.

- 30. The method of Claim 22 wherein said fatty acid composition is enriched for C12 fatty acids.
- 31. The method of Claim 22 wherein said fatty acid composition is enriched for at least one medium chain fatty acid and at least one other medium chain fatty acid is decreased.
- 32. The method of Claim 31 wherein said enriched fatty of acid is C12 and said decreased fatty acid is C14.

48	96	144	192	240	2 8 8	336	384
GGC	AAG Lys	GGT Gly	CAC His	GGG G1y	TCA	GCT	ACT
C CCG	c TCC u Ser	G GGT t Gly	G GGT s Gly	c ATG n Met	c TAT n Tyr	r GCC a Ala	A GGC Y GlY
GAT CCC Asp Pro	CGC CTC Arg Leu	GGA ATG Gly Met	G AAG u Lys	ACA AAC Thr Asn	CCA AAC Pro Asn	T GCT.	GCT GGA Ala Gly
GTG GAT Val Asp	GAC CGC Asp Arg		ATC GAG Ile Glu	ATT ACA Ile Thr	GGC CCA Gly Pro	TTC CAT Phe His	ATT GCT Ile Ala
CTA	GCC Ala	GGA Gly	CTT	GCC	ATG	TGC	ATG
GAA	GGT Gly	GTC Val	TCT	TAT Tyr	CTC	TAC	CTT
CTA	CTC Leu	CTG	CAG Gln	CCC	GGT	AAC Asn	GAT ASP
GCT Ala	GAT ASP	GTG Val	GTT Val	ATC Ile	TTT Phe	TCC	GCT Ala
GCC AAla	GCC FAIA	GGA Gly	GGG G1Y	TTC Phe	GAA Glu	ACT Thr	r GAG
GCG Ala	A CGA	GCC J Ala	GAC Asp	TTC Phe	ATC Ile	r GCC s Ala	r GGT J Gly
GTG A Val	GCA Ala	s AGA 1 Arg	TCT Ser	CCT Pro	GCT Ala	A TGT	CGT JArg
GCG Ala	r TCG	GAG Glu	TTC Phe	ACC Thr	CTC	r GCA : Ala	cGC Arg
ACC Thr	s AAT y Asn	AAG Lys	r GTC : Val	A ATC	CTG	ACT	r ATC s Ile
Ser	: AGG	GAC ASP	3 ACT 1 Thr	s aaa g Lys	r GCC Ala	r TCC	r CAT n His
AGC	TGC	ATC	CTG	CGG	TCT Ser	ATT Ile	AAT

FIGURE 1 1 OF 4

432	480	528	576	624	672	720	768
AGG	TGG Trp	TTG	ATT Ile	ACT Thr	AGC Ser	GCT Ala	ATC Ile
TGC Cys	CCC	GTG Val	ATT Ile	ATG Met	AGT	AAT Asn	GCC
GCT	AGG Arg	GGA GTG Gly Val	CCG	CAC	GAG Glu	ATA Ile	AAT Asn
GTG Val	TCT	GCT Ala	GCA	TAT Tyr	ATT Ile	TAC Tyr	ATA
TTT Phe	GCC	GGT	GGA Gly	GCT	TGC Cys	AAT Asn	GAG Glu
66C 61y	ACT Thr	GAA Glu	CGA	GAT	TCT	GTC Val	GCC
GGA Gly	CAG	GGT	AGA Arg	TGT Cys	TCT	GAG Glu	$r_1 = r_2$
TTG	CCG	ATG Met	ATG Met	AAC	GTC Val	GAA Glu	GAT CTC ASP Let FIGURE 2 OF 4
666 G1y	GAC	GTG Val	GCA Ala	ATC Ile	GGT	CCT	GGG G1y
ATT Ile	GAT Asp	TTT Phe	CAT	GCA Ala	CTT	TCA	GCT Ala
ATT CCA Ile Pro	AAC	GGT Gly	GAA Glu	GGT Gly	GGT Gly	GTC Val	CTA
ATT Ile	AGG	GAT Asp	TTG	GGA G1y	GAT	GGC Gly	ACT Thr
ATC Ile	CAA Gln	CGT Arg	AGC Ser	TTG	GCT	GCT	TCT Ser
GCA	TCT Ser	GAC	GAG Glu	TAT	AGG	GAT Asp	ACT
GCC	TTG	AAA Lys	ATG Met	GAG Glu	CCA	GAA Glu	GCG Ala
GAG Glu	GCT Ala	GAT Asp	GTG Val	GCA Ala	GAT	CTT Leu	CAT His

FIGURE 1 3 OF 4

816	864	912	096	1008	1056	1116
ATT AAT GCA ACT AAG	GGT CTT GAA GCT ATA	CAT CCC AGC ATT AAT	ACT GTT GCC AAC AAG	AAT TCA TTC GGA TTT	TTC AAG CCA TGATTA	TACGGATTAT GGACTTGCAG AGTAATTTCC 1 GTTGTCCGTC AAACCCATTT AGGATACTGT 1
Ile Asn Ala Thr Lys	Gly Leu Glu Ala Ile	His Pro Ser Ile Asn	Thr Val Ala Asn Lys	Asn Ser Phe Gly Phe	Phe Lys Pro	
AAC ACA AAG GAT ATC AAA	TGT CTT GGA GCA TCT GGA	ATA AAC ACC GGC TGG CTT	CCA TCG GTG GAG TTC GAC	GTT AAC GTT GCG ATC TCG	TCA GTC GTG GCT TTC TCG GCT T	TCATTGAGAA
Asn Thr Lys Asp Ile Lys	Cys Leu Gly Ala Ser Gly	Ile Asn Thr Gly Trp Leu	Pro Ser Val Glu Phe Asp	Val Asn Val Ala Ile Ser	Ser Val Val Ala Phe Ser Ala P	
AAG AAG GTT TTC AAG	TCA ATG ATC GGA CAC	GCG ACT ATT AAG GGA	CAA TTC AAT CCT GAG	AAG CAG CAA CAC GAA	GGA GGC CAC AAC TC	CCCATTTCAC AAGGTACTTG
Lys Lys Val Phe Lys	Ser Met Ile Gly His	Ala Thr Ile Lys Gly	Gln Phe Asn Pro Glu	Lys Gln Gln His Glu		CCATGTTTGT CGGAAGAGCA

FIGURE 1 4 OF 4

1236 1296 1348 TCTATGTAAT AAAACTAAGG ATTATTAATT TCCCTTTTAA TCCTGTCTCC AGTTTGAGCA TGAAATTATA TTTATTTTAT CTTAGAAAGG TCAAATAAGA TTTTGTTTTA CCTCTGTAAA ACTITIGITI GIAITGGAAA GGAAGIGCCG ICTCAAAAAA AAAAAAAA AA

Sequence Range: 1 to 1704

40 GTG Val>		GCA Ala>		TCT Ser>	0	GAC Asp>	240	cgg Arg>	CTC Leu>		GAA Glu>
GNG GTG XXX Val		AAT TCG Asn Ser	140	GAC	190	ATC Ile		ATC Ile	AGG		GCC GGG AAG AAG GCT CTC GAA Ala Gly Lys Lys Ala Leu Glu>
ACC	90	AAT Asn	•	GTC Val		TTA		CAG	30 AGG Arg	330	GCT
30 TCC Ser		AGG Arg		GAC		AGC	230	GGC GGC CAG Gly Gly Gln	280 GAC AGG ASP Arg		AAG Lvs
TGG AGC Trp Ser		GGC TGC Gly Cys	130	TCC	180	ATC Ile		GGC	AAC		AAG
	80		, 	GGC		GGG G1y		TTC	AAG Lys	320	GGG
20 AGC Ser		CCG		TTC		GGC GAG AGC Gly Glu Ser	220	AGG Arg	270 GGG AAG Gly Lys	(T)	GCC
AAA Lys		CCC		GTA Val	170	GAG	2	ACC	GAC		GTC Val
AAC	7.0	GAT Asp	120	TCC		GGC Gly		CCC	ATC Ile	0	ATT Ile
10 AAA GGG AAC Lys Gly Asn	•	GTG Val		GTC		TCC		TTC Phe	260 TAC	310	CGC TAC TGC ATT GTC Arg Tyr Cys Ile Val
		CTA		CTC Leu	160	CTC	210	AAG Lys	260 GGA TAC Gly Tyr		TAC
ACT		GAA Glu	110	ATG GGC CTC Met Gly Leu	1(CTC		TCC	ACG		CGC
CTC	09	CTA	••			GAA AAG Glu Lys		GCT Ala	50 GCG Ala	300	CTC
ACC Thr		GCT Ala		GGC G1y		GAA Glu	200	GAC GCT Asp Ala	250 AAC GCG Asn Ala		TGC
AAA TTA Lys Leu		GCG GCC Ala Ala	100	CGA GCC Arg Ala	150	TAC	(4	TTC	TTC		SAC GAT TGC CTC Asp Asp Cys Leu
AAA Lys	20	GCG Ala	7 (CGA		\mathtt{TAT}		CGC Arg	GGA Gly	90	GAC

FIGURE 2

	GAT AAG GAG AGA Asp Lys Glu Arg>	430	TCT Ser>	480	CCG Pro>	GCC Ala>		TGT Cys>		CGA Arg>	0	ATT Ile>
380	GAG Glu	4	TTC		TCC	CTT			620		670	GCA ATC Ala Ile
` ,	AAG Lys		GTC		ATC Ile	20 CTG Leu	570	ACT GCA Thr Ala	9	ATC CGC Ile Arg		GCA
			ACC	470	AAG ATC Lys Ile	520 GCT CTG Ala Leu		TCA		CAT		GCT Ala
370	ATT Ile	420	CTA	7.	CGG Arg	TCT		ATT	0	AAT Asn	4	GAG Glu
'n	AAG Lys		GGC G1y		GGT CAC Gly His	GGG G1y	260	TCG	610	GCC		ACT
	TCC		$_{\rm GLY}^{\rm GGT}$	09	GGT Gly	510 ATG Met	u)	TAT		GCT		GGA ACT Gly Thr
	CTC	410	ATG Met	4(AAA Lys	AAC Asn		AAC Asn		GCC Ala	029	GGA Gly
360	AGC	•	GGT Gly		GAG Glu	ACA	20	CCA	*	TAT	Φ.	GCT
	GAA Glu		ACT		ATC Ile	500 ATT Ile	550	GGC Gly		TTT Phe		ATT Ile
	GGT G1y	400	GGA Gly	450	CTC	GCC Ala		ATG		TGC	0	ATG Met
350	GGC G1y	4	GTT Val		AAT Asn	TAT Tyr		CTG	590	TAC	640	CTC
•	CTC		CTA		CAG Gln	90 CCC Pro	540	$_{\rm GGT}$	L)	AAC Asn		GAC
	GAT Asp		GTG Val	440	GTT Val	490 ATT CCC Ile Pro		TTG		TCC		GCT
340	TCC	390	GGA GTG Gly Val	7'	GGG Gly	TTC Phe		GAT	0	ACT	630	GAG Glu
m	AAT		GCT		GAC	TTT Phe	30		580	GCT		GGC Gly
										•		

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720	CAA AGG Gln Arg>	GAT Asp>		TTG Leu>		GAA TAT TTG GGA Glu Tyr Leu Gly>	01	GAT Asp>	960	GGG Gly>	ACT Thr>	
	CAA	CGT Arg		GAG AGC Glu Ser	860	TTG GGA Leu Gly	910	GCT GAT Ala Asp		GCT	TCC	
	TTA TCT Leu Ser	760 AAG GAC Lys Asp	810	GAG AGC Glu Ser	ω	TAT		AGG		GAA GAT Glu Asp	ACT Thr	
710				ATG		GAA Glu		GAT CCA AGG Asp Pro Arg	950		1000 GCG ACT Ala Thr	
	GCT Ala	GAT		TTG GTT Leu Val	0.0	GCA Ala	900	GAT Asp	O1	CTG	CAT	i
	TGC AGG Cys Arg	750 CCG TGG Pro Trp	800	TTG	850	ATT		ACT		AGT	GCT	
00	TGC Cys	750 CCG Pro	w	GTA Val		ATT Ile		ATG ACT Met Thr	01	AGC	990 AAT Asn	
70	GCC Ala	AGG		GGA G1y		CCG	890	CAT	940	GAG Glu	ATA Ile	
	GTT Val	TCA	790	GCT Ala	840	GCG Ala	ω	TAT		ATT Ile	TAC	
	GGA TTC Gly Phe	740 GCC	75	GGGG		GGA Gly		GCT		TGC	980 AAT Asn	
069	GGA Gly	ACT		GAA Glu		CGA Arg	0 %	GAT Asp	930	TCT Ser	980 GTC AAT Val Asn	
	GGA Gly	CAG		GGC Gly	830	AAA CGA Lys Arg	880	TGT		TCC Ser	GAG	
	GGG TTA Gly Leu	730 GAC CCT ASP Pro	780	ATG Met	ω	ATG Met		AAT TGT Asn Cys		GTC Val		
089	GGG Gly	73 GAC ASP		GTG Val		GCA Ala		GTC Val	920	GGT	970 CCT GAA Pro Glu	
•	ATT Ile	GAT		TTT Phe	0	CAT His	870	GCA Ala	O	CTT Leu	TCA	
	CCA	AAT	70	$\texttt{GGT}\\ \texttt{G1} \texttt{Y}$	820	GAA Glu		GGT Gly		GGG G1y	GTC Val	

FIGURE 3

	TTC AAG Phe Lys>		ATC GGA CAC Ile Gly His>	0.0	AAG GGA Lys Gly>	1200	GAG Glu>	GAA Glu>		TCA Ser>		GCA
	TTC AAG Phe Lys	1100	GGA CAC Gly His	1150	AAG GGA Lys Gly	T	CCC			AAC Asn	1340	AAT
1050	GTT Val	1	ATC Ile		ACA ATT Thr Ile		TTC AAT Phe Asn	1240 CAG CAA CAT Gln Gln His	1290	GGC CAC AAC Gly His Asn	13	TGA TTA CTC GGT TCA AAT GCA
•	AAG Lys		ATG Met		ACA Thr	1190	TTC	1240 CAG CAA Gln Gln	П			GGT
	ATC AAG AAG GTT Ile Lys Lys Val	90	AAG TCG Lys Ser	1140	GCG ACA ATT Ala Thr Ile	11	CAA Gln	AAG Lys		TTC GGA Phe Gly	0	CTC
1040	ATC	1090	AAG Lys	, ,	ATT		AAC Asn	AAG AAG Lys Lys	1280	TTC GGA Phe Gly	1330	TTA
1(GCC Ala		ACT		GCC Ala	081	ATA Ile	1230 3 AAC 1 Asn	12	GGA Gly		TGA
	AAT		GCA Ala	130	GAA Glu	118	AGC	GCC Ala		TTC Phe		CCA
30	ATA Ile	1080	AAT Asn	11	CTT Leu		CCC	GTT Val	0,	TCA	1320	AAG Lys
1030	GAG Glu	``	ATC Ile		GGT Gly		CAT	1220 C ACA P Thr	127	AAT TCA Asn Ser	П	TTC
	GCC		ACA Thr	0	TCA GGG Ser Gly	1170	CTT Leu	1220 GAC ACA ASP Thr		TCA		GCC
	CTT Leu	1070	ATC Ile	1120	TCA	V 1	TGG Trp	TTC		ATC Ile	1310	TCA
1020	GAT Asp	1(GAA Glu		GCA Ala		GGC G1y	1210 GTG GAA Val Glu	1260	GCT Ala	4	TTC
	GGG G1y		AAG Lys		GGA Gly	1160	ACC Thr	1210 GTG G Val G	 1	GTT Val		GCT
	GCT Ala	0.9	ACC	1110	CTT	11	ACC Thr	TCA		AAT Asn	0	GTA Val
10	CTT	1060	AAC	, (TGT		ATA Ile	CCA	20	GTG Val	1300	GTT Val

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AATTTGTTGC TGAGACAGTG AGCTTCAACT TGCAGAGCAA TTTTTTACAT GCCTTGTCGT TATCTGTTTG CCCTTGTCAA TGGCATTTAA GATAAGCTTA TAAAAAAAA AAAAAAAA AAAACTCGAG CGGAAGAGCG TAATACCGGG ATAGTTCCTT GATAGTTCAT TTAGGATGTT TTACTGCAAT TTTCTTTTG GGGGGGCCCG GTACCCAATT CGCCCTATAG TGAGTCGTAT GACAATTCAC TGTCCGTCGG AATCGAAGAT TATTTCCATT CTAATCCAGT CTCCGNCGAG TTTGAGAATC TATTAGAAAG AACGAGGCAA GATTTTGTTT CATGTTTGTG TTTGTATTAC

FIGURE 2 5/5

09	GCGGTGGCGG CCGCTCTAGA ACTAGTGGAT	120	GGTCGGCTCA GCTCAGGTGT	TGT ACG TGG Cys Thr Trp		CGT TCC Arg Ser	0	CTC TCC Leu Ser	310	CCT TGC Pro Cys	360	TTC GGA Phe Gly
20	AGA A	110	rca g		210	GAC CCA CGT Asp Pro Arg	260	ACT (Thr I	(-)	GAT (Asp B		CTC 1 Leu E
	CTCT	V 1	יכפפכי	160 T TTC O Phe	(4	GAC		AGG Arg		CTC	350	TCC
	ວນນ		GGT	C CCT r Pro		AAC Asn		CGC CGG Arg Arg	300	CAA TGC Gln Cys	m	GCT
40	ອອວອ	100	TTCTTACTTG	G TCC a Ser	200	TCC GAC Ser Asp	250		.,	CAA Gln		GGA TTC Gly Phe
	GGTG		CTTA	150 T GCG 1 Ala	2			CGT		TTC		GGA Gly
0		0		GTT.		TCA		TCC	0	ACC Thr	340	AAC Asn
30	CCAC	90	SAGT	C ATG s Met		ACT	240	CTC	290	TCC		
	ACTAAAGGGA ACAAAAGCTG GAGCTCCACC		GGCACGAGTT	140 TCT TGC Ser Cys	190	CCC		CGC		GGA Gly		GGG GAT Gly Asp
20	TG G	80				ATG		CTC		CGC	330	CTC
	AAGC		GAAT	C GCT r Ala		TGC	230	AAG CGG Lys Arg	280	CTC	(.,	TTC
	ACAA		GCAG	0 G ACC a Thr	180	GCT GCA Ala Ala	7	AAG Lys		TCC		CGC
10	GGA	70	CCCCGGGCT GCAGGAATTC	130 G GCG t Ala				CAC		TGC	320	CAG CAA Gln Gln
	AAAG		ದರ್ಧ	A ATG Met		GTA Val		TCC	270	CAT His	33	CAG CAA Gln Gln
	ACT		ညည	TCCA	170	CTC	220	CTT	• •	TCC		AAC Asn

FIGURE 3 1/6

ACT		GAA Glu		GTG Val		TAC	009	AAC Asn	TCT		GAC
GGC CGC Gly Arg	•	GCA CAG Ala Gln	200	GTA GTT Val Val	550	GTT Val	W	ATA GAG AAC Ile Glu Asn	AAG Lys		GAG AGG ATG GAC Glu Arg Met Asp
	450		2(GTA Val		GAT Asp		ATA Ile	ATC Ile	069	AGG Arg
400 CTC Leu	•			CGA		GAC CCC Asp Pro	290	GAG Glu	640 GAG Glu	•	GAG AGG Glu Arg
CTG AGG Leu Arg		CAA		CAA AGG Gln Arg	540	GAC	5.5	AGT	GGA G1y		
	440	ATG Met	490		3 7	CAT His		ATA Ile	GCC	0 8	TTC
390 GGC CAC Gly His	4	GCT		ACC AAG Thr Lys		GGC		AGT GGC	630 ATT Ile	680	CCA AAG TTC TCC Pro Lys Phe Ser
		GTG Val		ACC	530	CTA GGC Leu Gly	580	AGT	AGA Arg		CCA
CGC		ATG GCT Met Ala	480	GCT	Ŋ	CCT		ATA Ile	ACG Thr		GCC Ala
380 TCA AAT Ser Asn	430	ATG Met	•	CCT		ACT		GGA Gly	20 CCC Pro	670	GTG Val
		GTC Val		AAA Lys		GTG Val	570	GAC	620 TTT C		TGG GTG Trp Val
CGT		GAG Glu	470	AAG Lys	520	GTG Val	u,	CTA	CAG Gln		GGC Gly
CTT Leu	420	$\frac{GGG}{G1Y}$	4	AAT Asn		GGC G1y		CTC	TCT Ser	* 099	GAT Asp
370 CCT Pro		TCC		ACA Thr		ATG Met	260	AAT Asn	610 TGC Cys	U	ACA GAT Thr Asp
AAG Lys		CAT		TCC	510	GGT	2(AAC Asn	GAC Asp		TCC Ser
TCC	410	TCC	460	GTC	u ,	ACA		TAC	TTC	650	TTT Phe

FIGURE 3 2 OF 6

0	GAT Asp		TGT	840	GAT	TGT Cys		GAC		ACA		GAA Glu
740	GCA	790	AAG TGT Lys Cys	ω	AGC	TTT Phe			0 8		1030	
	TTA		AGA Arg		TTC	CCC	930	CTT GCA ATG Leu Ala Met	980	TGT GCA Cys Ala	₩.	AAA GGC Lys Gly
	GCA		AAT AAA AGA Asn Lys Arg	830	GTA	880 AGT Ser	O1			GCC		ATC Ile
730	GGC AAG AAA GCA Gly Lys Lys Ala	780		œ	AAG Lys	ATC Ile		ATT Ile		TCA ACT Ser Thr	1020	CAC ATA ATC His Ile Ile
•	AAG Lys	•	CTC		GGT ATG Gly Met	AAG AAG Lys Lys	920	GCT Ala	970	TCA	70	CAC
			GAG Glu		$\texttt{GGT}\\ \texttt{G1y}$	870 AAG Lys	6	TCC		ATA Ile		AAC Asn
720	GCA Ala	770	AAA Lys	820	66C 61y	TAT TYT		GGA Gly		TCG	0	GCG Ala
7	ACT	7.	ATG		TTG	TCA		ATG Met	096 *	TAT Tyr	1010	GCT
	CTG		GCG Ala		GGA G1y	860 AGG ACT Arg Thr	910	AAT Asn	O1	AAC Asn		AAT Asn
0	* ATG Met		GAA GAT Glu Asp	810	TCC	86 AGG Arg		ACA Thr		CCT		CTG
710	TAC	760	GAA Glu	•	GGC G1y	CTG		ACC Thr	950	ATG GGC Met Gly	1000	ATA Ile
	CTT		ACT Thr		ATT Ile	GCT	*	TCT Ser	9,	ATG Met	C	TGT Cys
	ATG Met		GGA ATC Gly Ile	800	CTC	850 GAA Glu	O1	$\mathbf{T}\mathbf{T}\mathbf{T}$		TGG Trp		TTC
700	TTC	750		8	GTT Val	ATT Ile		CCT Pro		GGA Gly	066	T AAC r Asn
•	AAG Lys	• -	GGT G1Y		GGA Gly	TCC	890	GTA Val	940	TTG	O)	AGT

FIGURE 3 3 OF 6

1080	GTT Val	AAT Asn		TTT Phe		CAT		AGT	1320	GCT	TCG
7	CCT	AAT		GAT GGA Asp Gly	20	TTA GAG Leu Glu	1270	GGT GGG Gly Gly	13	GGA GCT Gly Ala	GTC Val
	TTA	AGG	1170	GAT Asp	1220	TTA Leu	V 1	GGT Gly		GAA	GGA GTC Gly Val
0 /	GTT Val	1120 CAG Gln	त्न	CGT		GAG Glu		CTA	0	CAC CCT GAA His Pro Glu	
1070	GCC GTT Ala Val	TCA Ser		AAT Asn		GAG Glu	1260	$ ext{TTT}$	1310	CAC CCT His Pro	1360 GCT CAG TCC Ala Gln Ser
	GAT GCG Asp Ala	TTG	20	GAC AGT Asp Ser	1210	CTT Leu	12	GAA Glu		CCT Pro	GCT
	GAT	1110 3A GCT 9 Ala	1160		V -1	CTT Leu		GCG			rh 🗃
1060	TCG	11 CGA Arg		TGG Trp		TTA	0	\mathtt{TAT}	1300	ACC GAG Thr Glu	1350 GCC TTC Ala Let
•	GGC	TGC		CCA	1200	GTT Val	1250	ATT TAT (Ile Tyr		ATG	AAG Lys
	GGT Gly	1100 GTA GCA Val Ala	1150	AGA Arg	12	GGA Gly				CAC	
1050	TGT Cys	1100 GTA G Val A	V- V	TCG		GCT			1290		1340 ATA GAG Ile Glu
H	CTT Leu	TTC		GCT	06	GGA G1y	1240	AGA GGT GCA Arg Gly Ala	12	GCC TAC Ala Tyr	TGC
	ATG	GGT Gly	1140	AAA Lys	1190	GAA GGA Glu Gly	4			GAC	CTC
40	* GAC ATG ASP Met	1090 TTG GGA Leu Gly	[ACC Thr		GGA Gly		AAA Lys	0	TGC	1330 ATC Ile
1040		TTG		CCT		ATG Met	1230	GCA AAG AAA Ala Lys Lys	1280	ACT TGC Thr Cys	1330 GTG ATC Val Ile
	GCA	GGT G1y	1130	GAC	1180	GTG Val	12	GCA Ala		TTC	GGT Gly

FIGURE 3 4 OF 6

	GCT		AAC Asn		CTT	1560	AGG	GGC Gly		GTC Val		TCC
	TCC ACT CCT Ser Thr Pro	0.5	GGC CAA Gly Gln	1510	CTT CTT Leu Leu	7	ATA AGG Ile Arg	GAC GAA GGC Asp Glu Gly		AAG Lys	0	TCA
1410	ACT Thr	1460	GGC Gly	Н	CAC His		GCA	GAC Asp	1650	CTG	1700	AAC
17	TCC		TTC Phe			0			16	AAA CTG AAG Lys Leu Lys		CAT
	CAT GCA ACT TCC ACT CCT GCT His Ala Thr Ser Thr Pro Ala		TGT Cys	1500	ATC GGT Ile Gly	1550	GTT CAG Val Gln			AAG AAG GAG AAA CTG AAG GTC Lys Lys Glu Lys Leu Lys Val		TTC GGC GGC CAT AAC Phe Gly Gly His Asn
00	GCA Ala	1450	GCC CAC TGT Ala His Cys	15	ATG				0	AAG Lys	1690	GGC G
1400	CAT His		GCC Ala		TCG Ser		GCA GTA Ala Val	1590 T TTG GAA n Leu Glu	1640	AAG AAG Lys Lys	H	TTC
	GCG Ala		CTC	0	_	1540	GTT Val	15 AAT Asn		CCT		GGG
	ATA AAT Ile Asn	1440	GCT	1490	ACC AAA Thr Lys	-	GCA Ala	ATT Ile		GGC G1y	1680	
1390		16	CAA GCT Gln Ala		TCC		GAA Glu	AAT Asn	1630		16	TCA TTT Ser Phe
• •	TAC		TAC		AAT Asn	1530		1580 CCA AAT Pro Asn		CTC GTC Leu Val		AAT
	AAT Asn	0 8	AAG GAA Lys Glu	1480	GTG AAT Val Asn	15	GGC GTA Gly Val	CAT		CTG	0	TCC
1380	GAC GTA Asp Val	1430	AAG Lys	П	AGA		GGT	ATC Ile	1620		1670	TTG
``			ATC Ile		CTG	0	GCT		16	GCA AAA Ala Lys		GGT Gly
	GAA Glu		GAT Asp	1470	GAG Glu	1520	GGA GCT Gly Ala	1570 GGA TGG Gly Trp		GAT Asp		GTC
1370	AGG	1420	GGA	14	AGT		GGA G1y	ACA Thr	1610	GTG Val	1660	AAG

FIGURE 3 5 OF 6

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01/1	1/20	1/30	0 T / 40	1750	1760	
ATA CTA TI	ATA CTA TTT GCC CCC TGC AAC TAG A Ile Leu Phe Ala Pro Cys Asn ***	GC AAC TAG A	A AAAGAGTCTG		TGGAAGCCGA GAGTCTTTGA	
1770	1780	1790	1800	1810	1820	
GAACTCATGC	BAACTCATGC ACGTTAGTAG	CTTCTTATGC	CTCTGAAACC GAGATAGACC		GGCTACTCGA	
1830	1840	1850	1860	1870	1880	
GGGGATGCCA	AAGATACTCC	TTGCCGGTAT	TGGTGTTAAG	AGATCACTGC	TTGTCCCTTT	
1890	1900	1910	1920	1930	1940	
TATTTTCTTC		TTCTTTTGAG AGCTTTAACC	GAGGTAGTCG '	TATTTCGAG CTTTTCGAAT	CTTTTCGAAT	
1950	1960	1970	1980	1990	2000	
ACATGTTCGT	TATCGGATCA	ATGTGTTTCT	TCTAAGATCA	TTTGTAATGC	ATATTTTGAA	
2010	2020	2030	2040			
AAACCACATC	TCAGTATGCA	AAATAAAAA	AAACCACATC TCAGTATGCA AAATAAAAA AAAAAAAA AAAAAA	AAAAA		

1921
1 to
Range:
Sequence

0 *	TTCGAGCCCT GCCATGACTA CTACACCTCC	120	ACCACCGGCA GGCACCGGAG GCTCAATCGA	180	AATGGCTGTG GCTCTGCAAC CTGCACAGGA AGTTACCACA	GGA ATG Gly Met>		AT AAT sn Asn>		TTT GAT TGT Phe Asp Cys>	370	TTC TCC ACA Phe Ser Thr>
	CE		J B		AG		0	TAC AAT Tyr Asn	320			TTC TCC Phe Ser
20	CTA	110	GAG	170	GGA	220 G ACT 1 Thr	270					
	ATGA		ACCG		SACA	22 GTG Val		TTC		ACC		TCT
	CCC		GGC7		CTG(GTT Val		GAT GTT Asp Val	310	GAG Glu	360	GAG ATC AAG TCT Glu Ile Lys Ser
40	CCCT	100	CGCA	160	CAAC	GTA Val	260		, m			ATC Ile
	CGAG		CACC		TCTG	210 CGA Arg	•	CCT		GAG Glu		GAG Glu
0		0			GG	CGG		GAC Asp		AGT Ser	350	GGA G1y
30	CGGCACGAGG TCACCTCTTA CCTCGCCTGC	90	GCCCATCCGC	150	CTGTG	CAG Gln	250	CAT His	300	ATA Ile	m	GCT GGA Ala Gly
	CTCG		CCCA		ATGG(200 ATC AAA Ile Lys	2	GGC Gly		GGC Gly		ACG AGA ATT Thr Arg Ile
20	ra c	80		140		ATC Ile		CTA Leu		AGT Ser	0	AGA Arg
	CTCT		ATCC	ř	3GAG(AGT		CCT	290	GGA ACG Gly Thr	340	
	TCAC		TCGGATCCAG		CCGGGGAGGC	190 AAG CCA Lys Pro	240	ACT Thr				CCT Pro
10	AGG	70		130		AAG Lys		GTG Val		GAT Asp		TTT Phe
	CACG		GCATCCTTGT	` '	GCTTCCCCTT	AAG Lys		GGT GTG Gly Val	30	CTG CTT Leu Leu	330	CAA Gln
	CGG		GCA'		GCT	AAG Lys	230	$_{\rm G1Y}^{\rm GGT}$	280	CTG		GCT

FIGURE 4

FIGURE 2/6

420	ATG Met>	460 AAT GGT GGA ATC Asn Gly Gly Ile>		CTC Leu>		GCC ATT GAA Ala Ile Glu>	0	TTC Phe>	099	TGG Trp>	TTT Phe>
	TTC	GGA		GTT Val	260	GCC ATT Ala Ile	610	CCT		GGA TGG Gly Trp	AAC Asn
	AAG Lys	460 AAT GGT Asn Gly	510	GGA GTT Gly Val	u,	GCC		GTA Val			
410	GAC AAG ASP LYS	4(AAT Asn		TGC		GAT		\mathtt{TGT}	029	ATG GAC TTG Met Asp Leu	700 TGT GCA ACG AGT Cys Ala Thr Ser
7	ATG Met	TTA ACA Leu Thr			0 9	AAT	*	TTT Phe	φ		GCA Ala
	AAG AGG ATG Lys Arg Met		200	AAA AGA AAA Lys Arg Lys	550	TTC Phe		CCC		CTT GCA Leu Ala	TGT Cys
00	AAG Lys	450 GCA Ala	u i	AAA Lys		GTA Val		AAT Asn	O	CTT Leu	690 GCT Ala
40	TCC	AAA Lys		GAT Asp		AAG Lys	290	ATG Met	64	ATG	ACT
	CTC	AAG Lys	0 6	CTA	540	ATG	LC)	AAG Lys		GCT	TCT Ser
	AAG Lys	440 GCC GGC Ala Gly	490	GAG Glu		GGA G1y		AAG Lys		TCA	680 ATA Ile
390	CCG	GCC Ala		AAA Lys		GGT Gly	0 8	\mathtt{TAT}	630	GGA Gly	680 TCG ATA Ser Ile
	GCC Ala	ACT		ATG Met	530	ATG Met	580	TCA		ATG	TAC
	GTG Val	430 G CTG t Leu	480	GTG Val		GCA Ala		ATT Ile		AAT Asn	70 AAC Asn
380	TGG GTG Trp Val	43 ATG Met		GAT Asp		TCA		AGG	620	ACA Thr	670 CCC AAC Pro Asn
• •	GGT Gly	TAC		GAA Glu	0	GGC Gly	570	CTA	φ	ACC Thr	GGC
	GAT	CTT	470	ACC	520	ATT Ile		GCC		GCT	ATG

SUBSTITUTE SHEET (RULE 26)

	GTG Val>		GGA Gly>	850	ACT Thr>	* 006	GGG Gly>	AAA Lys>		TGC Cys>		ATT Ile>
	GAT Asp	800	GGT ATG Gly Met	8	CCT		ATG Met	940 CAT GCA AAG His Ala Lys		TTC ACT Phe Thr	1040	GTG Val
750	GCA Ala	~	GGT Gly		GAC		GTT Val	940 T GCA s Ala	066	TTC	10	GGA GTG Gly Val
	GAA Glu		ATT Ile		AAT GCC Asn Ala	890	GGA TTT GTT Gly Phe Val	94 CAT His		AGT		GCT Ala
	ATC AGA GGC GAA GCA GAT Ile Arg Gly Glu Ala Asp	790	CCT Pro	840		w		TTA GAG Leu Glu		GGA Gly	0	CCT GAT GGA GCT Pro Asp Gly Ala
740	AGA Arg	75	ATA Ile		AGA		GAT	TTA	980	CTA GGT GGA Leu Gly Gly	1030	GAT Asp
•	ATC Ile		ATC Ile		CAG	880	CGT Arg	930 GAG Glu	Oi	CTA		CCT
	ATA Ile		GTA Val	830	TCA	88	AAT Asn	GAG Glu		TTT Phe		CAC His
30	CAC	780	GCG Ala	w	TTG		AGT	CTA	0,	GAA Glu	1020	CCT
73	AAC		GAT Asp		GCT		GAC	920 CTA Leu	970	GCA GAA Ala Glu	П	GAG Glu
	GCG Ala		TCA	820	CGA Arg	870	TGG Trp	CTA Leu		TAC		ACC Thr
	GCT Ala	770	GGC Gly	83	TGC		CCA	GTG Val		ATT Ile	1010	ATG Met
720	AAT Asn	•	GGG G1Y		GCA		AGA Arg	.0 GGA Gly	096	ACT Thr	10	CAC ATG His Met
	CTG		TGC		GTT Val	860	TCA	910 GCT G Ala G		GCG Ala		TAC
	ATC Ile	760	CTT Leu	810	TTT Phe	w	GCT	GGA Gly		GGT Gly	00	GCC Ala
710	TGT	7(ATG Met		GGT G1y		AAA Lys	GAA Glu	950	AGA	1000	GAT GCC Asp Ala

3/6

	•											
06	GAA GAC Glu Asp>	1140	GAT ATC Asp Ile>	TTA Leu>		GCC Ala>		GGG TGG Gly Trp>	0	GAT ACC Asp Thr>	1380	GGT Gly>
1090	GAA Glu		GAT Asp	GAG Glu		CTC GGA GCA GCC Leu Gly Ala Ala	880	ACT GGG Thr Gly	1330	GAT ACC Asp Thr	~	GTC Val
	AGG		GCT GGA Ala Gly	1180 AAC AAC Asn Asn	1230	GGA Gly	1280	ACT Thr		GTG		AAG
	TCT	1130	CCA GCT Pro Ala		•			AGG Arg			1370	ATT Ile
1080	GGA GTC Gly Val	सं	CCA	CAA Gln		CTT	0,	GCA ATA AGG Ala Ile Arg	1320	GA G1	13	CTG AAC ATT AAG Leu Asn Ile Lys
•			ACT	GGC Gly	1220	CAC His	1270	GCA Ala	-	GAT Asp		CTG
		50	TCC	1170 TTC Phe	12	$_{\rm GGT}$		CAG Gln		CCA	0	AGA Arg
1070	CAG Gln	1120	ACA Thr	TGT		ATT Ile		GTT Val	1310	AAC Asn	1360	GAG Glu
H	GCT		GCC Ala	CAC	0 1	ATG	1260	GTA Val	13	GAA Glu		AAG Lys
	TTG		CAT	1160 CTT ATC Leu Ile	1210	TCA ATG Ser Met	Н	TCA		TTG		AAG Lys
20	AAG GCT Lys Ala	1110	AAT GCA Asn Ala	11 CTT Leu		AAA Lys		GTT Val	0	AAT	1350	
1060	AAG (Lys		AAT	GCT		ACC Thr	1250		1300	ATT AAT Ile Asn	[GGC CCT Gly Pro
	GAG Glu		ATA Ile	1150 TAC CAA TYr Gln	1200	TCT	12	GAA GCA Glu Ala		AAT Asn		GTG Val
	ATA Ile	1100	TAC	1150 TAC C. TYF G	7	AAT Asn		GTG Val		CCG Pro	1340	CTC
1050	TGC	H	AAT Asn	GAG		GTG Val	0	$\texttt{GGT}\\ \texttt{Gl}_{Y}$	1290	CAT His	13	TTG
• •	CTC		GTA Val	AAA Lys	1190	At to	1240	GGT GGT Gly Gly	⊣	ATC Ile		AAA Lys

FIGURE 4

() () () ()			1540	ATG	1600	CAG	1660	AAT	1720	4CA	1780	SCG	1840	3AG
CTC TTC Leu Phe>	1480	CAAA	Ħ	CATGCCCATG	1(GGCGACACAG	7(TTTCTGAAAT	1.7	GAAGAGAACA	1.7	TTTATCGCCG	18	ATCATTGGAG
1420 TCG TCC ATA Ser Ser Ile	1470	ATTCTACTCA ATCTATCAAA	1530	CGTCTCTAGA C	1590	GAGTACTCAT G	1650	TCCCATTTTT T	1710	AGTCAGTGAA G	1770	TGCTCTCTAT T	1830	TTTTCTCTTG A
1410 GGG CAC AAC Gly His Asn	1460	TGGA	1520	тасстсстта	1580	ATGACGGATT	1640	CTATTCATTA	1700	CGTTTCATCG	1760	CCCTTTGTTT	1820	GACTGGTTTG
1400 GGG TTT GGT (Gly Phe Gly (1450	TAG GGCGTTT CATGTG	1510	AGCATGTTGG	1570	AGTCGGAACC	1630	TGTTAGAGCA	1690	TACTTTCGAG	1750	TAACCATTTG	1810	AAAACTAGAC
TCA TTC Ser Phe	1440		1500	TGAGGACTCC	1560	CGGGAGCTGT	1620	TTGCTAGAAT	1680	ACGGTAGTTG	1740	GGGCACGTAG	1800	TAAAATTTGT
1390 TTG TCT AAT Leu Ser Asn	1430	GCC CCT TAC AAC Ala Pro Tyr Asn	1490	GCTGAAGTTT	1550	AGTTTTGTGT	1610	GATATACTCC	1670	CTCCCTCCTT	1730	AAGCTAACTC	1790	TTTTGTGGGT

IGURE 4 5/6

1850 1860 1870 1880 1890
ATGTATGGCC ATATTTGCCT TTCATTGATG ATAAAAAAA AAAAAAAA AAAA
1910 1920

1900

AAAAAAAA AAAAAAA A

FIGURE 4

09	120	169	217	265	313	361	409	457	505
CTGGTACGCC TGCAGGTACC GGTCCGGAAT TCCCGGGTCG ACCCCACGCGT CCGTCTTCCC	ACTCCGATCG TTCTTCTTCC ACCGCATCTC TTCTCTTCTC	CGCCGCC ATG CAT TCC CTC CAG TCA CCC TCC CTT CGG GCC TCC CCG CTC Met His Ser Leu Gln Ser Pro Ser Leu Arg Ala Ser Pro Leu 1	GAC CCC TTC CGC CCC AAA TCA TCC ACC GTC CGC CCC CTC CAC CGA GCA Asp Pro Phe Arg Pro Lys Ser Ser Thr Val Arg Pro Leu His Arg Ala 25	TCA ATT CCC AAC GTC CGG GCC GCT TCC CCC ACC GTC TCC GCT CCC AAG Ser Ile Pro Asn Val Arg Ala Ala Ser Pro Thr Val Ser Ala Pro Lys 35	CGC GAG ACC CCC AAG AAG CGC GTC GTG ATC ACC GGA ATG GGC CTT Arg Glu Thr Asp Pro Lys Lys Arg Val Val Ile Thr Gly Met Gly Leu 55	GTC TCC GTT TTC GGC TCC GAC GTC GAT GCG TAC TAC GAC AAG CTC CTG Val Ser Val Phe Gly Ser Asp Val Asp Ala Tyr Tyr Asp Lys Leu Leu 65	TCA GGC GAG ATC GGC CCA ATC GAC CGC TTC GAC GCC TCC AAG Ser Gly Glu Ser Gly Ile Gly Pro Ile Asp Arg Phe Asp Ala Ser Lys 85	TTC CCC ACC AGG TTC GGC GGC CAG ATT CGT GGC TTC AAC TCC ATG GGA Phe Pro Thr Arg Phe Gly Gln Ile Arg Gly Phe Asn Ser Met Gly 100	TAC ATT GAC GGC AAA AAC GAC AGG CGG CTT GAT GAT TGC CTT CGC TAC Tyr Ile Asp Gly Lys Asn Asp Arg Arg Leu Asp Asp Cys Leu Arg Tyr 115

FIGURE .

553	601	649	697	745	793	841	688
GCC	666 G1y	CTT Leu	GCC Ala 190	ATG Met	TGC Cys	ATG Met	GGC Gly
GGT G1y	GTT Val	TCT Ser	TAT Tyr	CTG Leu 205	TAC	CTT	GGA Gly
CTC Leu 140	CTG	CAA	CCC Pro	GGT	AAC Asn 220	GAT Asp	TTG
GAT Asp	GTG Val 155	GTT Val	ATC Ile	CTC	TCC Ser	GCT Ala 235	GGG G1y
GCC Ala	GGA Gly	GGG G1y 170	TTC Phe	GAA Glu	ACT Thr	GAG Glu	ATT Ile 250
GAC	GCC Ala	GAC	TTC Phe 185	ATT Ile	GCC	$_{\rm G1Y}^{\rm GGT}$	CCA
GAG Glu	AGA Arg	TCT	CCT	GCT Ala 200	$ ext{TGT}$	CGT	ATT Ile
CTT Leu 135	GAG Glu	TTC Phe	ACC Thr	CTC	GCA Ala 215	CGC	ATC Ile
TCT Ser	AAG Lys 150	GTC Val	ATC Ile	CTG	ACT Thr	ATC Ile 230	GCA
AAG Lys	GAC	ACT Thr 165	AAA Lys	GCC Ala	TCC	CAT	GCC Ala 245
AAG Lys	ATC Ile	CTG	CGG Arg 180	TCT Ser	ATT Ile	AAT Asn	GAG Glu
666 G1y	AAG Lys	$_{\rm G1Y}^{\rm GGT}$	CAC	GGG G1y 195	TCA	GCT Ala	ACT
GCC Ala 130	TCC Ser	GGT	GGT Gly	ATG Met	TAT Tyr 210	GCT	GGC Gly
GTC Val	CTC Leu 145	ATG Met	AAG Lys	AAC Asn	AAC Asn	GCT Ala 225	GGA G1y
ATT Ile	CGC Arg	GGA G1y 160	GAG Glu	ACA Thr	CCA	CAT	GCT Ala 240
TGC	GAC	ACA Thr	ATC Ile 175	ATT Ile	GGC	TTC	ATT Ile

FIGURE 5

937	985	1033	1081	1129	1176	1224	1272
ACT Thr 270	GAA Glu	CGA Arg	GAT Asp	TCT Ser	GTC Val 350	GCC Ala	AAA Lys
CAG Gln	GGT G1y 285	AAA Lys	TGT Cys	TCC Ser	GAG Glu	CTC Leu 365	ATC Ile
CCT Pro	ATG Met	ATG Met 300	AAC Asn	GTC Val	GAA Glu	GAT Asp	GAT ASP 380
GAC Asp	GTG Val	GCA Ala	ATC Ile 315	$_{\rm GLY}^{\rm GGT}$	CCT	GGG G1y	AAG Lys
GAT ASP	TTT	CAT His	GCA Ala	CTC Leu 330	TCA Ser	GCT	ACA Thr
AAC Asn 265	GGT Gly	GAA Glu	GGT Gly	${\tt GGT} \\ {\tt G1y}$	GTC Val 345	CTA	AAC Asn
AGG Arg	GAT Asp 280	TTG	GGA G1y	GAT	GGC Gly	ACT Thr 360	AAG Lys
CAA	CGT	AGC Ser 295	TTG	GCT Ala	GCT Ala	TCT Ser	TTC Phe 375
TCT	GAC	GAG Glu	TAT Tyr 310	AGG Arg	GAT	ACT Thr	GTT Val
CTG	AAA Lys	CTG	GAG Glu	CCA Pro 325	GAA Glu	GCG Ala	AAG Lys
GCT Ala 260	GAT	GTG Val	GCA Ala	GAC	CTT Leu 340	CAT His	AAG Lys
AGG Arg	TGG Trp 275	TTG	ATT Ile	ACT Thr	AGC Ser	GCT Ala 355	ATC Ile
TGC Cys	CCC	GTG Val 290	ATT Ile	ATG Met	AGT Ser	AAT Asn	GCC Ala 370
GCT	AGG Arg	GGA Gly	CCT Pro 305	CAC	GAG Glu	ATA Ile	AAT
GTG Val	TCT	GCT Ala	GCA Ala	TAT TYT 320	ATT Ile	TAC	ATA Ile
TTT Phe 255	GCC	GGT Gly	GGA Gly	GCT	TGC Cys 335	AAT Asn	GAG Glu

FIGURE 3/4

IJ

1320	1368	1416	1464	1512	3 1569	1629	1689	
CAC TGT CTT GGA GCC TCT GGA His Cys Leu Gly Ala Ser Gly 395	GGA ATA AAC ACC GGC TGG CTT Gly Ile Asn Thr Gly Trp Leu 410	GAG CCA TCC GTG GAG TTC GAC Glu Pro Ser Val Glu Phe Asp 425	GAA GTT AAT GTT GCG ATC TCG Glu Val Asn Val Ala Ile Ser 440	TCA GTC GTG GCT TTC TCG GCT Ser Val Val Ala Phe Ser Ala 460	CACTTGTC ATTGAGAGTA CGGTTGTTCG	NAAAAGTA AGGATTATCA CTTTCCCTTC	ATATTTATTT TAAAAAAA AAAAAAGGGC	
GCA ACT AAG TCA ATG ATC GGA Ala Thr Lys Ser Met Ile Gly 385	GAA GCT ATA GCG ACT ATT AAG Glu Ala Ile Ala Thr Ile Lys 405	AGC ATT AAT CAA TTC AAT CCT Ser Ile Asn Gln Phe Asn Pro 420	GCC AAC AAG CAG CAA CAC Ala Asn Lys Lys Gln Gln His 435	TTT GGA TTC GGA GGC CAC AAC Phe Gly Phe Gly Gly His Asn 450	CCA TGA TTACC CATTTCACAA GGCACTTGTC Pro 465	AT TTAGGATACT GTTCTATGTA AAAAAAGTA	TCCAGTTTGA GAATGAAATT	
ATT AAT Ile Asn	GGT CTT Gly Leu 400	CAT CCC His Pro 415	ACT GTT Thr Val	AAT TCA Asn Ser	TTC AAG Phe Lys	TCAAACCCAT	TAATCCTGTC	

FIGURE 5

Sequence Range: 1 to 1802

09	TTATCTCCGC	10 CCT TCC Pro Ser		TCC Ser	210	CGT Arg		CGG		GTC Val	CTA Leu
	TTAT	110 TCC C	160	TCC	••	ATC Ile		AAG Lys		GAC	350 ATC AGC Ile Ser
20		CAC		CCC Pro		GTC		CCC AAG Pro Lys	300	F S	
	ACATTTCATT TCTTGCCTCG	CTC		TCC	200	CCC	250	CCC Pro	• •	GGC Gly	GGC G1y
	\mathbf{TCT}	100 TCC Ser	150	AAT Asn	2	CTC		GAC		TTC	AGC
40	CATT	CAA		CTC		AGC		TCC Ser	290	GTC Val	340 GAG Glu
	ATTT	ATG (CGC		GCC	240	GAG Glu	2	TCC	GGC Gly
0		Ö	140	TTC	190	CGC		CGC		GTC Val	TCC
30	CGAC	၁၅၁၁၅	H	CCC		CGT Arg		AAG Lys		CTC	330 CTC Leu
	CTTTCCGACC	ລອລລອລລອລລ 06		GAG Glu		CTC	230	CCC	280	GGC G1y	330 CTG CTC Leu Leu
20				CTC	180	CCC	7	GCC Ala		ATG Met	AAG Lys
	CGCGTCCGGG	80 CCGTCGTTCG	130	CCT	••	CGC		TCC		GGC Gly	320 TAC GAC Tyr Asp
	CGCG	CCGT		TCC		CTC		GCC Ala	270	ACC	32 TAC TYE
10				CCC	170	GCT	220	ACC Thr	•	ATC Ile	TAC
	GGTCGACCCA	70 CGCTCCTCCG	120	CGC	H	GCC		GCC Ala		GTC Val	GCC Ala
	GGT(CGC		CTC		GCC		GCT Ala	260	GTC Val	310 GAC ASP

FIGURE 6 1/5

	GGC CAG Gly Gln	450	CGG Arg		GCT		AAG Lys	GTC Val		ATC Ile	069	CTG
400	GGC CAG Gly Gln	4.	GAC CGG Asp Arg		AAG Lys		GAT Asp	90 ACT Thr	640	AAG Lys	9	GCG CTG Ala Leu
	GCC		AAC Asn		GGC AAG Gly Lys	540	ATT GAT Ile Asp	590 CTA A		CGG Arg		TCT
	ACC AGG TTC GCC Thr Arg Phe Ala	440	GGC AAG Gly Lys	490	GGC AAG Gly Lys	u,	AAG Lys	GGC Gly		CAC His	30	GGG G1y
390	AGG Arg	4	GAC GGC AAG Asp Gly Lys		GCC		TCC	GGT Gly	630	AAA GGT CAC Lys Gly His	680	ATG
					GTC Val	530	CTC	580 ATG Met	W	AAA Lys		ACA AAC Thr Asn
	CCC		ATC Ile	480	ATT GTC Ile Val	5.5	TCC	GGT		GAG Glu		ACA Thr
380	TTC	430	TAC	•	TGC		CAA Gln	ACC Thr	620	ATC Ile	019	ATT
ä	AAA Lys		66C		TAC		GGC Gly	570 GGA G1y	39	CTC		GCC
	TCC		ACG	470	CGC	520	GCC	GTT Val		AAT Asn		TAT
	GCT	420	GCG Ala	4,	CTC		CTC	CTA Leu		CAG Gln	099 *	CCA
370	GAC	•	AAC Asn		TGC		GAT Asp	560 A GTG Y Val	610	GTT Val	v	ATT Ile
	TTC		TTC		GAT Asp	510	GCC Ala	56 GGA G1Y		$\frac{GGG}{G1y}$		TTC Phe
	CGC	10	GGC Gly	460	GAC	u ,	GAC	GCC		GAC	0 9	TTT Phe
360	GAC	41	CGT		CTC		GAA	AGG	* 009	TCT	650	CCG
•	ATC Ile		ATC Ile		CGG	200	CTC	550 GAG Glu	•	TTC		TCC

FIGURE 6 2/5

FIGURE 3/5

	ACT		ATC	525	Ala		TCT	930	GAC		GAG Glu		TAT Tyr
	TCA ACT Ser Thr		CAT His	GCT	Ala	880	TTA	on .	AAG Lys		ATG Met		GCG CCG ATT ATT GCA GAA TAT Ala Pro Ile Ile Ala Glu Tyr
	TCG ATT Ser Ile	780	GCC AAT CAT Ala Asn His	8 GAG	07n		GCT		CCG TGG GAT AAG Pro Trp Asp Lys		TTG GTT Leu Val	1020	GCA Ala
730	TCG	•		ACT	Thr		TGC AGG Cys Arg	920	TGG	970	TTG	7(ATT Ile
	GGC CCA AAC TAT Gly Pro Asn Tyr		GCC	GGA	GIY GIY Thr	870		6			GTA		ATT Ile
	AAC Asn	770	GCT				GCC		AGG		GGA Gly	10	GCG CCG ATT Ala Pro Ile
720	CCA Pro	7	TAT	GCT	Ala		GTT Val		TCA	096 *	GCT	1010	GCG Ala
			TTT Phe	ATT	Ile	09	TTC	910	GCC Ala	•	GGG Gly		GGA G1y
	ATG Met		TGC	810 ATG	Leu Met	98	GGA		ACT		GAA Glu		CGG
710	CTG	760	TAC				GGA Gly		CAG Gln	950	GGT Gly	1000	ATG AAA Met Lys
7	GGT Gly		AAC		Asp		TTA	006	CCT	9	ATG Met	•	
	TTG		TCC	800 GAG GCT	Ala	850	GGT Gly		GAT		GTG Val		GCA Ala
	GAT Asp	750	ACT	8 GAG	GIn		ATT Ile		GAT Asp		TTT Phe	066	GAG CAT Glu His
700	ATC Ile		GCT		GIY		CCA	890	AGG AAT Arg Asn	940	GGC Gly	•,	GAG Glu
	GCC		TGT	CGA	Arg	840	ATT Ile	ά			GAT Asp		TTG
	CTT	740	GCA	790 CGC	Arg		GTC		CAA		CGT	980	AGC

SUBSTITUTE SHEET (RULE 26)

AGG		GAT Asp	1170	ACT		GTT Val		ATC Ile	ATT Ile		AAT
CA	1120	GAA Glu	11	GCG ACT Ala Thr		ATT AAG AAA GTT Ile Lys Lys Val		ATG		1360	CCC AGC ATT AAT CAA TTT AAT Pro Ser Ile Asn Gln Phe Asn
1070 GAT C ASP P	• •	CTC		CAT His		AAG Lys	1260	TCA	1310 GCA ACC Ala Thr		CAA
ACT		AGT	20	AAT GCT Asn Ala	1210	ATT Ile	H	AAG Lys	ATC Ile		AAT
ATG Met	1110	AGC	1160		\ \	GCC		ACT	GCC	1350	ATT
1060 TAT CAT Tyr His	 i	GAG Glu		ATA Ile		ATA AAT Ile Asn	20	AAT GCA Asn Ala	1300 ' GAA Glu	73	AGC
		ATT Ile		AAT TAC Asn Tyr	1200		1250	AAT	CTT		CCC
GCT	00	TGC	1150	AAT Asn	H	GAG Glu		ATC Ile	GGT	10	CAT
1050 T GAT S ASP	1100	TCG		GTC		GCC Ala		AAA Lys	1290 TCA GGA Ser Gly	1340	TGG CTT CAT Tro Leu His
7G CY		TCC		GAG	06	CTT	1240	ATC Ile	12 TCA Ser		TGG
AAC		GTC Val	1140	GAA	1190	GAT	``	GAA Glu	GCA Ala		GGC Glv
1040 GCA GTC Ala Val	1090	GGT G1y	÷ή	CCT		GGG G1y		AAG Lys	1280 CTT GGA Leu Gly	1330	ACC ACC Thr Thr
	•	CTT Leu		TCA		GCT Ala	1230	ACC Thr	1280 CTT G Leu G	• •	ACC
GGT Gly		GGG G1y	30	GGG GTC Gly Val	1180	CTT	13	AAC Asn	TGT Cys		ATA Ile
GGA G1y	1080	GAT Asp	1130	GGG	• •	ACT		AAG Lys	CAC	1320	AAG GGA ATA Lvs Glv Ile
1030 TTG Leu	J.	GCT		GCC		TCT	1220	TTC	1270 GGA G1Y	73	AAG
					•						

4/5

1410	s cag caa s Gln Gln		GGA GGG CAC Gly Gly His	1510	ATTCT ACTTGGTTCA	1570	TAAATGCCTT	1630	AGCCATTTAG	1690	CTCTGATTTA	1750	GTTATTTAAG		CT
1400	AAC AAA AAG CAG CAA Asn Lys Lys Gln Gln	1450	GGA TTT GGA GGG CAC Gly Phe Gly Gly His	1500	TGA ATTCT /	1560	AGCAATTTTT	1620	AGTTCCTCGA AGCCATTTAG	1680	TAAATCTAGT	1740	TGTTGTCAAT GTTATTTAAG	1800	ATCCAGCTTA
1390	ACT GTT GCC Thr Val Ala	1440	AAT TCT TTT Asn Ser Phe	1490	TTC AAG CCA Phe Lys Pro	1550	CAACTTGCAG	1610	GTCCTTTGAT	1670	ATTCCCATTT	1730	GTCATGTTTG	1790	GCTCTAGAGG ATCCAGCTTA
	GAC TTC AAC ASP ASP '	1430	GCT ATC TCG A	1480	TTC TCA GCT Phe Ser Ala 1	1540	GATAGGGCTT	1600	GAATAGGTCG	1660	ATCGAAGATG	1720	AAGATTTTGT	1780	AAGGGCGGCC
1380	TCG GTG Ser Val		AAC GTC Asn Val	1470	GTG GCA Val Ala	1530	CAGTTGCTGA	1590	CGTAATACCG	1650	TACTGTAATA	1710	AGACCAATGA	1770	ATAAAGCAAA AAAAAAAAA AAGGGCGGCC
1370	CCC GAG CCA Pro Glu Pro	1420	CAT GAA GTG His Glu Val	1460	AAC TCG GTT Asn Ser Val	1520	AAATGCACAC	1580	GTCGGAAGAG	1640	GATGATGTTT	1700	TGTATTAGAA	1760	ATAAAGCAAA

F160KE 0 5/5

Sequence Range: 1 to 2369

	CATAAAAGAG	120	TTACCATACC	180	ATCCTTTTCT	230 TCT TCC Ser Ser>	280	ATG TCT Met Ser>	330	TCT CCT Ser Pro>		CCA CTA Pro Leu>
20	CACGCGTCCG CATAAAAGAG	110	CTTCGATTCA TTACCATACC	170	CCCAAAGGGT	220 ATG CCT GCC GCC Met Pro Ala Ala	270	GCC GCC TGC Ala Ala Cys	320	TCC ATC TCC Ser Ile Ser	370	CAA TGC GCC Gln Cys Ala
40	CGGGTCGACC	100	CTCCTTTCAT	160	GGTCTTTCAT CCCAAAGGGT	220 CCTCCA ATG C	0.0	TGG CTC CTT Trp Leu Leu	310	CTT CCG CCT Leu Pro Pro	360	TCC
30	CCGGAATTCC	06	TGCGGCCACC	150	GCCTTTTCCG	210 CAGTCAGTTC	260	TGT ACG Cys Thr	O *	GAC CCT Asp Pro	350	CGC CGC CGG A Arg Arg I
20	AGGTACCGGT	80	ATCCATCGAA TGCGGCCACC	140	TCCATTTTCC	200 CTCAAAGGGT	250	TCC CCT CTC Ser Pro Leu	300	CAC CCC TCC His Pro Ser	340	CGC CTC TCC CG Arg Leu Ser Ar
10	GTACGCCTGC AGGTACCGGT	70	AGAGAGAGGG	130	ATTCCGCTGA	190 ATCCTATCTT	240	CTG CTC GCT Leu Leu Ala	290	ACC TCC TTC Thr Ser Phe	3	CGC CGA CGC Arg Arg

FIGURE 7

	CTC GTC Leu Val>	TCC Ser>	0 2	CGG Arg>	570	CTG Leu>		CAG Gln>		CAT His>	ATA Ile>
	CTC GTC Leu Val		520	CAC		GCT	•	AAA Lys			
420	ACC Thr	470 TAT ACA Tyr Thr		AGG Arg		GTG Val	0	ATC Ile	660	CTA GGC Leu Gly	AGT Ser
	CAT His	TAC		CGC AGG Arg Arg	260	GCC GTG Ala Val	610	AGT Ser		CCT	ACG
	TTC	460 CAT GAC His Asp	510	ACC	U)	ATG Met		AAG CCA Lys Pro		ACT	700 GAT GGA Asp Gly
410	AGT	46 CAT His		ACC		GCA Ala		AAG Lys	650	GTG ACT Val Thr	700 GAT GGA ASP Gly
7	TCC	TGC Cys		CGC	550	GAG Glu	600	AAG	w w	GTG Val	CTT
	GGA Gly	CCC	200	ATT Ile	55	AGG		AAG Lys		GGT Gly	CTG
400	CGC	450 GAG Glu	u,	CCC		TCC		ACA Thr	079	ATG Met	690 AAT Asn
4 (CTC	TTC		AGA		CCT Pro	590	ACC	9	GGA ATG Gly Met	AAT Asn
	GCC	TGC	490	TCC	540 *	TCC	u,	GTT Val		ACT Thr	TAC
	TCC	440 CTC GCC Leu Ala	49	GGA Gly		GCT Ala		GAA Glu		GTG Val	680 TTC Phe
390	TCC	CTC		TTC Phe	•	CGA Arg	30	CAG	630	GTT Val	GTT Val
	GCT	TAC		TTG	530	AAT Asn	58	GAA Glu		GTA Val	GAT Asp
	TCT	10 TCT Ser	480	TCC	ц	CTC		CCT		CGA Arg	70 CCT Pro
380	CCT	430 ACC TCT Thr Ser		GCA Ala		AGG		CAA	620	CGG	670 GAC C ASP P

150RE /

760	ATT GCT Ile Ala>	810	CTC Leu>		AAG Lys>		CTA Leu>	950 GGA ATG Gly Met>	0.0	AAG AAG Lys Lys>	1050	GCT Ala>
7			AAG Lys				GAG Glu	950 GGT GGA Gly Gly	1000	AAG AAG Lys Lys		TCA
	AGA Arg		CCG	850	ACC GCT GGC Thr Ala Gly	006	AAA Lys	GGT G1y		$\mathtt{T}\mathtt{A}\mathtt{T}$		GGA Gly
	CCT ACG AGA Pro Thr Arg	800	GTG GCC CCG Val Ala Pro	8	ACC		ATG Met	ATG		TCA Ser	1040	AAT ATG GGA TCA GCT Asn Met Gly Ser Ala
750	CCT	~			CTG		GTG Val	940 TCA GCA ATG Ser Ala Met	066	ATT Ile	10	AAT Asn
	TTT Phe		TGG		TAC ATG Tyr Met	890	GAA GAT GTG Glu Asp Val	940 TCA G		AGG		ACC ACA Thr Thr
	CAA Gln	06	\mathtt{GGT}	840	TAC	ω	GAA Glu	GGC G1y		CTA	0	ACC
740	GCT Ala	79(GAT Asp		CTA		ACC Thr	ATT Ile	086	GCC Ala	1030	GCT
•	TGT		ACA Thr		ATG Met	30	ATC Ile	930 CTC Leu	O	GAA Glu		TTC
	GAT		TCC	830	TTC	880	GGA G1y	GTT Val		ATT		CCT
730	TTT Phe	780	TTC	w	AAG Lys		GGT Gly	$\frac{GGA}{G1y}$	0	GCC Ala	1020	GTA Val
7.	ACC Thr		TCT		GAC Asp		GAT Asp	920 TGC Cys	970	GAT GCC Asp Ala	1 1	TGT GTA Cys Val
	GAG Glu		AAG Lys	820	ATG Met	870	ACA Thr	920 AAA TGC Lys Cys		AAT Asn		TTT Phe
	ATA Ile	770	ATC Ile	8	AGG Arg		TTA	AGA Arg		TTC Phe	1010	CCC
720	GAG Glu		GAG Glu		AAG Lys		GCA Ala	10 AAA Lys	960	GTA Val	10	AAT Asn
	AGC		GGA Gly		TCT	860	AAA Lys	910 GAT AJ ASP L		AAG Lys		ATG Met

FIGURE 7

	TCT Ser>		CAT His>	90 GAT GCG Asp Ala>	40	GCT TTG Ala Leu>	1290	AGT Ser>		CTA Leu>		GAA Glu>
	ATA Ile		AAC Asn	1190 A GAT I ASP	1240	GCT Ala		GAC Asp		CTA Leu		GCA Ala
0	TCG	1140	GCG	11 TCA Ser		CGA Arg		TGG Trp	0		1380	
1090	TAC TCG Tyr Ser	↔	GCT GCG Ala Ala	GGC Gly		TGC	1280	CCA TGG Pro Trp	1330	GTG CTA Val Leu	ᆏ	ATT TAC Ile Tyr
	AAC Asn		AAT Asn		1230	GCA Ala	12	AGA Arg		GGA Gly		ACT Thr
		1130	ATG Met	1180 TGC GGG Cys Gly	⊣	GTT Val		TCA		GCT Ala	1370	
1080	GGG CCC Gly Pro	₩.	ATA Ile	CTT Leu		TTT Phe	0	GCT	1320	GGA Gly	13	GGT GCG Gly Ala
П	ATG		TGT Cys	ATG	1220	$_{\rm GGT}$	1270	AAA Lys	₹-1	GAA Glu		AGA Arg
	TGG	0	rrr Phe	1170 GrG	12	GGA Gly		ACT Thr		666 61y	0.9	AAA Lys
1070	GGA Gly	1120	AAC TTT Asn Phe	1170 GAT GTG ASP Val		ATG Met		CCT	1310	ATG Met	1360	AAG AAA Lys Lys
1(TTG		AGT	GCA Ala	0,	GGT Gly	1260	GAC	+	GTT Val		GCA Ala
	GAC		ACG Thr	1160 GGC GAA Gly Glu	1210	ATT Ile	\	TCC		TTT		CATHis
20	GCA ATG Ala Met	1110	GCA	1160 GGC GAA Gly Glu		CCT		AAT	0.0	GGA Gly	1350	GAG Glu
1060	GCA		TGT	aga Arg		ATA	1250	CAG AGA Gln Arg	1300	GAT Asp	V -1	TTG
	CTT Leu		GCT Ala	ATC Ile	1200	ATC Ile				CGT Arg		GAG Glu
	ATG	1100	ACT (Thr	1150 ATA ATC Ile Ile	П	GTA Val		TCC		AAT Asn	1340	GAG Glu

SUBSTITUTE SHEET (RULE 26)

1430 ACC GAG CCT Thr Glu Pro>	0 8	TTG GCT Leu Ala>	1530	CAT GCC His Ala>		CAC His>		ATG Met>	GTA Val>	0 2	GAA Glu>
1430 ACC GAG CCT Thr Glu Pro	1480	TTG GCT Leu Ala	-	CAT His		ATC Ile		TCA	1670 GTT TCA Val Ser	1720	TTG GAA Leu Glu
14 ACC Thr		GCT		GCC Ala	0 /	CTT Leu	1620	ACC AAA Thr Lys	16' GTT' Val		AAT Asn
ATG Met		AAG Lys	1520	ATA AAT GCC Ile Asn Ala	1570	GCT CTT Ala Leu	C · 4	ACC Thr	GCA		ATT Ile
1420 TAC CAC TYr His	1470	ATA GAG AAG Ile Glu Lys	-	ATA Ile		CAA Gln		TCA	1660 GTG GAA Val Glu	1710	CCG AAT Pro Asn
14. TAC TYE	• •			\mathtt{TAC}		GAG TAC Glu Tyr	1610	AAT Asn		• •	
GAT GCC Asp Ala		CTC TGC Leu Cys	10	GTA AAT Val Asn	1560	GAG Glu	7	GTT Val	GGT GGT Gly Gly		CAT
GAT Asp	160	CTC	1510	GTA Val	` '	AAA Lys		AAA Lys	GGT Gly	1700	TGG ATC Trp Ile
1410 ACT TGC Thr Cys	14	ATT Ile		GAC		ATC Ile	00	GAG TTA Glu Leu	1650 GCA GCC Ala Ala	, H	
ACT		GTG Val		GAA Glu	1550	GGA GAT Gly Asp	1600	GAG	GCA Ala		GGG G1y
TTC	20	GCT GGA Ala Gly	1500	AGG Arg	₩.	GGA		AGA Arg	GGA G1y	06	AGG ACT Arg Thr
1400 GGG AGT Gly Ser	145		V 1	TCT		GCT		CAA AAC Gln Asn	1640 CTT CTC Leu Leu	1690	
		GGA Gly		GTC Val	40	CCG	1590		1 CTT Leu		ATA Ile
GGT Gly		GAT Asp	190	GGA G1y	1540	ACT Thr	• •	GGC	CAC		GCA
90 CTA Leu	1440	CCT	14	TCA		TCC		\mathtt{TTC}	30 GGT G1y	1680	CA G1
1390 TTT C	•	CAC		CAG		ACA	1580	TGT Cys	1630 ATT G Ile G	•	GTT Val

FIGURE 7 5/7

AAG AAG Lys Lys>		TTT GGT Phe Gly>	1870	GTTTCCGTGT	1930	GTTGGTAGCT	1990	GAACCATGAC	2050	GTAGAGCAAT	2110	GTTGTACTTT	2170	CACGTAGTAA
GTG GGT CCT Val Gly Pro	1810	TCA TTT GGG Ser Phe Gly	1860	ATC TAG GAC Ile ***>	1920	ACTCCAGCAT	1980		2040		2100	CCTTGCAATA	2160	
AA TTG CTC	1800	TCSe	1850	SC CCT TAC	1910	AGTTTTGAGG	1970	TGTGTCCGGA	2030	ACTCCTTGCT	2090	AAATCTCCCT	2150	AACAAAGCTG
GAT ACA Asp Thr	190	GTC GGT Val Gly	1840	CTC TTC Leu Phe	1900	CAAAGCTGA	1960	CATGAGTTT	2020	ACTTGATAT	2080	TTTTCTCTG	2140	GAAGAAGAG
GAA Glu		GTT Val	1830	TCG TCC Ser Ser	1890		1950		2010		2070	TCATATTTTT T	2130	ATCGAGTCAG TGAAGAAGAG AACAAAGCTG TTAACTCGGG
AAC CCA GAT Asn Pro Asp	17	GAG AGA CTG Glu Arg Leu	1820	GGG CAC AAC Gly His Asn	1880	GTGGAATTCT	1940	CCTTACGTCT	2000	GGATTGAGTA	2060	ATTCATTATC	2120	CGAGCTTTTC
	CCA GAT GAA GGC GTG GAT ACA AAA TTG CTC GTG GGT CCT AAG Pro Asp Glu Gly Val Asp Thr Lys Leu Leu Val Gly Pro Lys	CCA GAT GAA GGC GTG GAT ACA AAA TTG CTC GTG GGT CCT AAG Pro Asp Glu Gly Val Asp Thr Lys Leu Leu Val Gly Pro Lys 1780 1790 1800 1810	CCA GAT GAA GGC GTG GAT ACA AAA TTG CTC GTG GGT CCT AAG Pro Asp Glu Gly Val Asp Thr Lys Leu Val Gly Pro Lys AGA CTG AAC GTT AAG GTC GGT TTG TCT AAT TCA TTT GGG TTT AAG GTC GGT TTG TCT AAT TCA TTT GGG TTT AAG GTC GGT TTG TCT AAT TCA TTT GGG TTT AAG GTC GGT TTG TCT AAT TCA TTT GGG TTT AAG GTC GGT TTG AAT TCA TTT GGG TTT AAG GTC GGT TTG AAT TCA TTT GGG TTT AAG GTC GGT TTG AAT TCA TTT GGG TTT AAG GTC GGT TTG AAT TCA TTT GGG TTT AAG GTC GGT TTG AAT TCA TTT GGG TTT AAG GTC GGT TTG AAT TCA TTT GGG TTT AAG GTC GGT TTG AAT TCA TTT GGG TTT AAG GTC GGT TTG AAT TCA TTT GGG TTT AAG GTC GGT TTG AAT TCA TTT GGG TTT AAG GTC GGT TTG AAT TCA TTT GGG TTT AAG GTC GGT TTG AAT TCA TTT GGG TTT AAG GTC GGT TTG AAT TCA TTT GGG TTT AAG GTC GGT TTG AAT TCA TTT GGG TTT AAG GTC GGT TTT GGG TTT AAG GTC GGT TTG AAT TCA TTT GGG TTT AAG GTC GTT TTG TTT GGG TTT GGG TTT AAG GTC GGT TTG AAT TCA TTT GGG TTT AAG GTC GTT TTG TTT GGG TTT AAG GTC GTT TTG TTT GGG TTT GGG TTT GTG TTT GGG TTT AAG GTC GTT TTG TTT GGG TTT GTG TTT GGG TTT TTT GGG TTT TTT GGG TTT TTT GGG TTT TTT GTG TTT TTT GTG TTT TTT GTG TTT TTT GGG TTT TTT GTG TTT TTT GTG TTT TT	C CCA GAT GAA GGC GTG GAT ACA AAA TTG CTC GTG GGT CCT AAG AA n Pro Asp Glu Gly Val Asp Thr Lys Leu Leu Val Gly Pro Lys Ly 1780 1790 1810 g AGA CTG AAC GTT AAG GTC GGT TTG TCT AAT TCA TTT GGG TTT GG TTT GGG TT	C CCA GAT GAA GGC GTG GAT ACA AAA TTG CTC GTG GGT CCT AAG AAA TTG CTC GTG GTG TABLO Lys Lya Leu Asn Val Lys Val Gly Leu Ser Asn Ser Phe Gly Phe Gly AAC GTC ATA CTC TTC GCC CCT TAC ATC TAG GAC GTTTCC AAC AAA TTG CCC CCT TAC ATA TCA TTT GGG	C CCA GAT GAA GGC GTG GAT ACA AAA TTG CTC GTG GGT CCT AAG AA TTG CTC GTG GT TTG TA TTG TCT AAT TCA TTT GGG TTT GG TTT	C CCA GAT GAA GGC GTG GAT ACA AAA TTG CTC GTG GGT CCT AAG AAA TTG CTC GTG GGT CCT AAG AAA TTG CTC GTG GGT CCT AAG LY	C CCA GAT GAA GGC GTG GAT ACA AAA TTG CTC GTG GGT CCT AAG AAA TTG CTC GTG GGT CCT AAG AAA TTG CTC GTG GGT CCT AAG AAA TTG CTC GTG GGT CT AAG LYS	C CCA GAT GAA GGC GTG GAT ACA AAA TTG CTC GTG GGT CCT AAG AAA TTG CTC GTG GGT CCT AAG AAA TTG CTC GTG GGT CCT AAG Lys Lys Leu Leu Val Gly Pro Lys Lys Lys Leu Lys Leu Lys Val Gly Leu Ser Asn Ser Phe Gly Pre Gly Pre Gly Lys Aug GTC GGT TTG TCT AAT TCA TTT GGG TTT GG TTT GGG GAACCAT GTT GGG GAACCAT GTT GGG GAACCAT GTT GGG GAACCAT GAACCAT GTT GGG GAACCAT G	C CCA GAT GAA GGC GTG GAT ACA AAA TTG CTC GTG GGT CCT AAG AAA TTG CTC GGT GGT CCT AAG AAA TTG CTC GGT CTG GGT CTG GGT TTG TCT AAT TCA TTT GGG TTT GG TTT GG TTT GGG TT GGG TTT GGG TT GGG TTT GGG TT GGG TTT GGG TT GGG TTT GGG TTT GGG TT GGG	C CCA GAT GAA GGC GTG GAT ACA AAA TTG CTC GTG GGT CCT AAG AA DE CA AAB GTG GTG GTG CCT AAG AAB TTG CTC GTG GGT CCT AAG AAB TTG CTC GTG GTG GTG CCT AAG TTG TTG TCT AAT TCA TTT GGG TTT GGG TTT GTG TCT AAT TCA TTT GGG TTT GGG TTT GGG TTT GTG TCT AAT TCA TTT GGG TTT GGG TTT GGG TTT GTG TCT AAT TCA TTT GGG TTT GGG TTT GGG TTT GGG TTT GGG TTT GTG TCG TC	C CCA GAT GAA GGC GTG GAT ACA AAA TTG CTC GTG GGT CCT AAG AAA TTG CTC GTG GGT CCT AAG AAA TTG CTC GTG GGT CT AAG AAA TTG CTC GTG GT CT AAG CTC GTG TTG TCT AAT TCA TTT GG TT GG TTT GG TTT GG T	C CCA GAT GAA GGC GTG GAT ACA AAA TTG CTC GTG GGT CCT AAG AA TTG CTC ASP GLU Leu Val Gly Pro Lys Lys Lys Leu Leu Val Gly Pro Lys Lys Lys Leu Leu Val Gly Bro Lys Lys Lys Leu Leu Asn Val Lys Val Gly Leu Ser Asn Ser Phe Gly Phe Gl As Has Asn Ser Ser Ile Leu Phe Ala Pro Tyr Ile ***> G CAC AAC TCG ATC ATA CTC TTC GCC CCT TAC ATC TTG GAC GTTTCC ATC ATC ATC TTG GAC GTTTCC ATC ATC ATC ATC ATC ATC ATC ATC AT	C CCA GAT GAA GGC GTG GAT ACA AAA TTG CTC GTG GGT CCT AAG AA 1780 1780 1780 1780 1780 1810

SUBSTITUTE SHEET (RULE 26)

				CTCTAGAGG	2360 AGGGGGGCCG
AAAAAAAAA	AAAAAAAA AAAAAAAAA AAAAAAAAA	AAAAAAAAA	AAAAAAAAA	TGGAAATAAA AAAAAAAA AAAAAAAA	TGGAAATAAA
2350	2340	2330	2320	2310	2300
ATGTATGTTT	TTCTCATTGA TAATTGGGGR ATGTATGTTT	TTCTCATTGA	TTGGTTTGTT	CTGGTTTAGA	AACTAGAAGA
2290	2280	2270	2260	2250	2240
AAATTTGTAA	TGTGGTTTTA AAATTTGTAA	ATCACCGTTT	TCTCTATTTC	TTTGTTTTGC	CCATTTGCCC
2230	\$220 *	2210	2200	2190	2180

FIGURE 7

2374
to
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Range:
Sequence

* 09	CACACCAAAC	120	ACAGACAGAC	180	TCTTCGATTC	240	TCCCAAAGGG	300	CCTGCCGCCT	360	TCTACCTCCT	420	CTCTCCCGCC	480	CGCGGATCCA
20	GACGCCAACC CACACCAAAC	110	CATTGGCAGC AGACAGACAG ACAGACAGAC	170	CCTCCTTTCA	230	GGGTCTTTCA TCCCAAAGGG	290	TCAGTCAGTT CCCTCCAATG CCTGCCGCCT	350	CGCCTGCATG	410	TCTCCTCTCC TCGCCGACGC CTCTCCCGCC	470	CTTCTGCTTC CTCCGCCCTC CGCGGATCCA
40	ACGCGTCCGC	100	CATTGGCAGC	160	ATGCGGCCAC	220	CGCCTTTTCC	280	TCAGTCAGTT	340	GGCTCCTTGC	400	TCTCCTCTCC	460	CTTCTGCTTC
30	GGGTCGACCC	06	AGACGGACGC	150	GATCCATCGA	210	ATCCATTTTC	270	TCTCAAAGGG	330	CTCTGTACGT	390	CCGCCTTCCA	450	CTCCCAATGC GCCCCACTAC
20	CGGAATTCCC	80	TCTCTTCTCA AGACGGACGC	140	GAGAGAGAGG	200	CATTCCGCTG	260	TATCCTATCT	320	CGCTTCCCCT	380	CGACCCTCTT	440	
10	-A-CNTGGTC	10	TTCCTCAGCT	130	CCATAAAAGA	190	ATTACCATAC	250	TATCCTTTTC	310	CTTCCCTGCT	370	TCCACCCCTC	430	GCCGGATTCT

FIGURE 8 1/5

540	GACTACTATA	* 009	CGGAGGCTCA	* 099	ACAGGAAGTT	720	AATGGGTGTG	780	ATGGAACGAG	840	CCTACGAGAA TTGCTGGAGA	*	GGATGGACAA	096 *	GAATCACCGA	1020	CAGCAATGGG
530	GCCCTGCCAT	290	TTCGCACCAC CCGCAGGCAC CGGAGGCTCA	650	GCCGTGGCTC TGCAACCTGA ACAGGAAGTT	710	CGGCGAGTAG TTGTGACTGG AATGGGTGTG	770	AATCTGCTTG	830	CCTACGAGAA	890	CTCTCTAAGA	950	ACAGATGGTG	1010	ATGCGGAGTT CTCATTGGCT CAGCAATGGG
520	CCTGCTTCGA	580	TTCGCACCAC	640	GCCGTGGCTC	700	CGGCGAGTAG	160	TTTCTACAAT	820	TGCTCAATTT	880	GGCCCCGAAG	940	GAAAGCATTA	1000	ATGCGGAGTT
510	TCTTACCTCG	570	TCCAGACCCA	630	GGAGGCAATG	069	TATCAAACAG	750	ACCTGATGTT	810	CCTTTGATTG	870	ATGGTTGGGT	930	CTGCTGGCAA	066	ATAAAAGAAA
200	CCTCGTCACC	260	CTTGTTCGGA	620	CCCTTCCAGG	089	AGAAGCCAAG	740	TAGGCCATGA	800	GAGATAGAGA	860	TTCTCCACAG	920	TACATGCTGA	980	AAAGAGCTAG
490	GTTTCCATAC	550	CATCCGCATC	610	ATCGAGCTTC	019	ACCACAAAGA	730	GTGACTCCTC	790	TGGCATAAGC	850	GATCAAGTCT	910	GTTCATGCTA	970	AGATGTGATG

FIGURE 8 2/5

TGCGATGCCT ACCACATGAC		GAGTTTCACT	TTCTAGGTGG	TACGCAGAAT	TGCGACTATT
1500	1490	1480	1470	1460	1450
AGAAAAGAGG	GAGCATGCAA	AGAGGAGTTG	TGCTACTACT	GGAGCTGGAG	TATGGGGGAA
1440	1430	1420	1410	1400	1390
ATGGATTTGT	AGTAATCGTG	ACCATGGGAC	AAGCTTCAAG	GACCCTACTA	GAGAAATTCC
1380	1370	1360	1350	1340	1330
CTTTGTCCCA	GCATGCCGAG	AGGTTTTGTT	TTGGTATGGG	ATCATACCTA	AGATGCGGTA
1320	1310	1300	1290	1280	1270
GCGGGGGCTC	GTGATGCTTT	CGAAGCAGAT	TAATCAGAGG	GCGAACCATA	AATGAATGCT
1260	1250	1240	1230	1220	1210
ACTTTTGTAT	GCAACGAGTA	TACTGCTTGT	ACTCGATATC	GGGCCCAACT	GGGATGGATG
1200	1190	1180	1170	1160	1150
CAATGGACTT	GCTATGCTTG	TATGGGATCA	CTACCACAAA	GTACCTTTCG	TCCCTTTTGT
1140	1130	1120	1110	1100	1090
AGAAGATGAA	ATTTCATATA AGAAGATGAA	AGCCCTAAGG	ATGCCATTGA	GTATTCAATG	TGGAATGAAG
1080	1070	1060	1050	1040	1030

FIGURE 8 3/5

TTTGTGTCCG	GCCCATGAGT	CTCTAGACAT	CTCCTTACGT	ATGTTGGTAG	GGACTCCAGC
2040	2030	2020	2010	2000	1990
GAAGTTTTGA	TATCAAAGCT	CTACTCAACA	GTGTGGAATT	GACGTTTCGT	TTACATCTAG
1980	1970	1960	1950	1940	1930
TCTTCGCCCC	TCGTCCATAC	TGGGCACAAC	TTGGGTTTGG	TCTAATTCAT	GGTCGGTTTG
1920	1910	1900	1890	1880	1870
TGAACGTTAA	AAGGAGAGAC	GGGTCCTAAG	AATTGCTCGT	GTGGATACAA	AGATGAAGGC
1860	1850	1840	1830	1820	1810
TGGAAAACCC	AATATTAATT	GATCCATCCG	GGACTGGGTG	CAGGCAATAA	TTCAGTAGTT
1800	1790	1780	1770	1760	1750
TGGAAGCAGT	GCCGGTGGTG	TCTCGGAGCA	TTGGTCACCT	AAATCAATGA	TAATTCAACC
1740	1730	1720	1710	1700	1690
AGTTAAAAGT	CTGTTTCGGC CAAAACAGAG AGTTAAAAGT	CTGTTTCGGC	CTCTTATCCA	GAGTACCAAG	AGATATCAAA
1680	1670	1660	1650	1640	1630
CTCCGGCTGG	GCCACATCCA	AAATGCCCAT	TAAATTACAT	AGGGAAGACG	AGGAGTCTCT
1620	1610	1600	1590	1580	1570
TGGCTCAGTC	TCTCTGCATA GAGAAGGCTT	TCTCTGCATA	CTGGAGTGAT	CCTGATGGAG	CGAGCCTCAC
1560	1550	1540	1530	1520	1510

FIGURE 8

		ATCC	2370 GCTCTAGAGG	2360 AAGGGCGGCC	2350 AAAAAAAAA
TTTTCTCAAA	GATTGGTTTG	GACTGGTTTA	AAAACTAGAA	TAAAATTTGT	TTTGTGGTTT
2340	2330	2320	2310	2300	2290
TCATCACCGT	GCTCTCTATT	CCTTTGTTTT	AACCATTTGC	GGCACGTAGT	TGTTAACTCG
2280	2270	2260	2250	2240	2230
AGAACAAAGC	AGTGAAGAAG AGAACAAAGC	TCATCGAGTC	TTCGAGCTTT	TAGTTGTACT	CTCCTTGCAA
2220	2210	2200	2190	2180	2170
TGAAATCTCC	TTTTTTCTC	TCTCATATTT	ATATTCATTA	TGGTAGAGCA	CTAGAATTGT
2160	2150	2140	2130	2120	2110
ATACTCCTTG	GACACTTGAT	TACTCATGGC	ACGGATTGAG	CGGAACCATG	GAGCTTTAGT
2100	2090	2080	2070	2060	2050

TGUKE 8

Sequence Range: 1 to 1580

	GGG G1y>	100	TCG Ser>	150	GTG Val>		GAT Asp>		TCT Ser>	AAA Lys>	01	CGC Arg>
20	H	1(CAT His		AGG		GGT Gly		ATT GGA Ile Gly	290 CTT GCT Leu Ala	340	ATC Ile
	AAT GCA Asn Ala		CAG Gln		TCC AAA Ser Lys	190	TTG	240	ATT Ile	CTT		GGG G1y
			ACT Thr	140	TCC	1.9	TCT		TTA Leu	GAT Asp		CGA ACG GGG ATC CGC Arg Thr Gly Ile Arg
40	ATG GCG Met Ala	90	GCA	\	GTC Val		CAG Gln		GGA TGC AAA TTA Gly Cys Lys Leu	280 AAT GAT Asn Asp	330	CGA Arg
			AGG Arg		TTT Phe		GAC AGG Asp Arg	230	TGC	280 AAT G Asn A		GTC Val
	GCTGGG		AGA Arg	130	GAG Glu	180	GAC	()	GGA Gly	TCA Ser		ACT Thr
0		80	CTG	Ή	TCG		TCT		AGA Arg	GTC Val	320	ATT Ile
30	rcgT'		GCC Ala		TCC		GAT Asp	220	AGT	270 CAA Gln	(.,	TGG Trp
	GAGTTTCGTT		CCT		TCT	170	CAG Gln	23	GTG Val	CTT		GAA Glu
		70	GTT Val	120	GGA Gly	• •	GTT Val		CTT	GCT	310	GAT Asp
•	ATTCAAGAGA	•	TCA		CGT		GCC Ala		AGG Arg	260 CCA Pro	31	AAT Asn
	ATTC		TCT		TCT	160	AGT Ser	210	CCG	2 ATA Ile		ACC Thr
			GGT Gly	110	TCG	1(TGT		TCG	GCT		GAC Asp
	CCTGAATCGG	6 09	CTG		TCA		TGC		CGC Arg	50 TCT Ser	300	GTC Val
	CCT		TTT Phe		ATT Ile		TTT Phe	200	TCT	25 GGT G1Y		ATT Ile

FIGURE 9

390	TCA Ser>		GAC GCA AAT GAT Asp Ala Asn Asp>		GGC Gly>	AAG AAT CCT TTG	580	TTG GGT TTA GTC Leu Gly Leu Val>	630	GTG Val>		GGA Gly>
	GCA Ala		GAC GCA AAT Asp Ala Asn		rrc Phe	530 CCT	25.	TTA GTC Leu Val		CTA GTG Leu Val		CGG Arg
	TTA Leu	430	GCA	480	CTT	AAT	TOW TO THE TOWN	GGT Gly		ATT Ile	0,	GAT Asp
380	ACA AAT TTA Thr Asn Leu	4	GAC		GAC Asp	AAG	s √⊓	TTG	620	AAT Asn	029	ACC Thr
V-1			GTA		ACC CCT GAG Thr Pro Glu	TGC AAA AAG AAT CCT TTG	570	GTG TTG GGT Val Leu Gly	9	TTT AAC AAT ATT Phe Asn Asn Ile		GAC TGG ACC GAT CGG Asp Trp Thr Asp Arg
	CTT Leu		GCA CAG GTA Ala Gln Val	470	ACC CCT GAG Thr Pro Glu	520 GGC TGC AAA	ر ک	TTT Phe		TTT Phe		GAC Asp
0 /	AGT	420	GCA Ala	7	ACC Thr	66C	ζ. 15	GGA Gly	0	GGT	* 099	GTT Val
37(GAT Asp		ATG Met		TCT	CTT	260	AGT	610	GGG		TAT
	AAA Lys		GAG Glu	0.5	ACT Thr	510 GCA	A 4 4 4 4 4 4 4 4 4 4 4 4 4 4 4 4 4 4 4	TGC		GGT Gly		CGG Arg
	GGT G1y	410	CTA	460	TGT Cys	AAA	ς Σ	GCA Ala		AGA Arg	029	TCT Ser
360	TCA	4.	GCT		ATG Met	TCG	0 Set	GCT Ala	* 009	ATT Ile	v	CTT
	CTC		AAA Lys		TTG	500 CAG ATA	250	ACC GCT Thr Ala		CAC		TCT
	GTT Val	400	AGG	450	GTT Val	CAG	5	ATT Ile		TGC	0 1	GAT ASP
350	AGG GTT Arg Val	4 (GCA		ATG Met	CCT	7	GAC	290	GCT	640	GCT Ala
•	CGA Arg		GCA Ala		GAT Asp	90 GCT	540	TAC Tyr	u,	GCT		GGT
	AAC		GAG Glu	440	GTG	AGT G	מעד	TCT		TCA		ATT Ile

IGURE

	CAG TCA Gln Ser>	770 CAT AGC GAT His Ser Asp>	820	AAA GAA GAT GAA GTT Lys Glu Asp Glu Val>	870	ATC AGA GAT TTT CCA CCA AGG Ile Arg Asp Phe Pro Pro Arg>		CGC Arg>		AAG Lys>	GCA Ala>
	CAG	770 AGC	œ	GAA GTT Glu Val		CCA		TTC CGC Phe Arg		GGA Gly	1010 T CAG S Gln
720	GTG Val	CAT His		GAT Asp		CCA	910	GTA Val	096	CTT Leu	1C CAT His
		TTG		AAA GAA GAT Lys Glu Asp	860	TTT	91	GGT AAA GAG GTA Gly Lys Glu Val		GCA	1010 TGG TTG CTG CTT CAT CAG Trp Leu Leu Leu His Gln
	GGA GCT GTA GTG Gly Ala Val Val	760 TTT GAT TTG Phe Asp Leu	810	AAA Lys	w	GAT Asp		AAC GGT AAA Asn Gly Lys		TCA	CTG Leu
710	GCT	7(TTT Phe		ATC Ile		AGA Arg		$_{\rm G1y}^{\rm GGT}$	950	ATC GAA TCA Ile Glu Ser	1000 TTG C Leu L
	GGA Gly	GCT Ala		GCA Ala	0.9	ATC Ile	006	AAC	ס	ATC Ile	TGG Trp
	GCT	TTT Phe	800	GCT	850	TCC		ATG		TCA	GAC Asp
700	GCT Ala	750 CTC Leu	w	AAA Lys		GGG Gly		CAA Gln	0.1	CAG Gln	990 ATC Ile
7(GAT Asp	GGG G1y		CTA		AAT Asn	068	ATC Ile	940	CCT	AAC Asn
	GGA G1y	GAT	790	CAT	840	CAT	ω	TGC		GTG Val	TCC Ser
	rrr Phe	740 GAA Glu	7	AGG		GGA Gly		TCT		TCT Ser	980 GGA G1Y
069	CTC	GAG Glu		CAA Gln		CTG	°0	\mathtt{TAC}	930	CGC Arg	9 AAT Asn
	ATT Ile	GCT		GGG	830	GCC Ala	880	TCA		${ m TGC}$	CTT
	TGT	30 GAT ASP	780	GAT Asp	ω	AAA Lys		TCT		GCT	70 GGT G1Y
089	ACA	730 TGT G Cys A		GGA Gly		GAT		CGT	920	TTT Phe	97 GCC Ala

FIGURE 9

46/66

AAT CCC ATC ATC ACA CCA ACA CCA ACA CCA C														
T CAG AGG ATC ATT GAT GCA GTA GCA T CAG AGG ATC ATT GAT GCA GTA GCA 1070 1080 1070 1080 1080 1090 1090 1100 1130 1130 1130 1220 1280 1230 1330 1340 1350 17TTTATGG AGCAAAC ACGAAGCAAT CTTA GCA 1330 1340 1350 1140 1150	1060		1110	GCG Ala		GTG Val		ACA	1260	CACTGCAGCT	1320	AAGAAGTCAG	1380	TCGTTCCCCT
T CAG AGG ATC ATT GAT GCA GTA GCA T CAG AGG ATC ATT GAT GCA GTA GCA 1070 1080 1070 1080 1080 1090 1090 1100 1130 1130 1130 1220 1280 1230 1330 1340 1350 17TTTATGG AGCAAAC ACGAAGCAAT CTTA GCA 1330 1340 1350 1140 1150	020	CTA GAG GTT Leu Glu Val	1100		1150		1200		1250	GCCGAGCCAG	1310	CCANAAAAAG	1370	
TOZO TOAG AGG ATC ATT GAT GCA GTA G I Gln Arg Ile Ile Asp Ala Val A 1070 1080 1120 1130 C ATT CCC TCA AAC TTG GCA A I Arg Ile Ile Ser Asn Leu Ala A I 1120 I 1130 C ATT CCC TTG GCA CTA GAC GAA G I Ile Pro Leu Ala Leu Asp Glu A I Ile Pro Leu Ala Leu Asp Glu A I Ile Pro Leu Ala Leu Asp Glu A I Ile Pro Leu Ala Ile Ala Thr Ala G G GGT CAC GTG ATT GCA ACC GCA G G GGT CAC GTG ATT GCA ACC GCA G G GGT CAC GTG ATT ATC AGG TGG GGA T Y Ser Ala Ile Ile Arg Trp Gly * I TCT GCT ATT ATC AGG TGG GGA T Y Ser Ala Ile Ile Ala Thr Ala G CTCTCAAA CCGATGTTTC ACGAAATTTT I 330 I 1340 I 1350		ACA	1090		1140	GTG	1190		1240	^	1300	3CTTCCATGA	1360	CTTCATCACA
T CAG AGG ATC ATT GA n Gln Arg Ile Ile As 1070 108 A CGA ATT ATC TCA AA u Arg Ile Ile Ser As 1120 C ATT CCC TTG GCA CT r Ile Pro Leu Ala Le 1170 1220 G GGT CAC GTG ATT GC o Gly His Val Ile Al 210 1220 T TCT GCT ATT ATC AG y Ser Ala Ile Ile Ar 1270 1280 TTTTATG AGCAAGCAAC	104	GCA GTA Ala Val		TTG GCA Leu Ala	130	GAC GAA Asp Glu	1180	ACC GCA Thr Ala	1230	TGG GGA Trp Gly	1290		1350	
AAT CAG AGG ASD 1070 GAA CGA ATT GGL ATG CGG GGT CAC GGT AGG AGG AGG AGG AGG AGG AGG AGG AGG	1030	ATT Ile	1080	TCA		GCA	170	ATT Ile	1220		1280	CGATGTTTC A	1340	
(-1	1020	CAG AGG Gln Arg	1070	CGA ATT Arg Ile	112	ATT CCC Ile Pro	1160	GGT CAC Gly His	1210	TCT GCT Ser Ala	1270	TCCTCTCAAA C	1330	TCTTTTATGG A

IGURE 9

				1580	1570 1580
AAAAAAAAA	TTTGCTAAAA AAAAAAAAA AAAAAAAA	TTTGCTAAAA	ATGTTTATAT	CATAAACATC ATGTTTATAT	GAGATGACAG
1560	1550	1540	1530	1520	1510
CGGGACATTG	CATTTTGTCT	GCTTTTACTT	TAATTGTTCA	GTTTCTTGTT	TAAGTTATTT
1500	1490	1480	1470	1460	1450
TTGTCCCCAA	ATAGTTTCTT	TACAATACCC	TTGCTGACAA	TTTGATGATT	TTTCCATTAG
1440	1430	1420	1410	1400	1390

IGURE 9

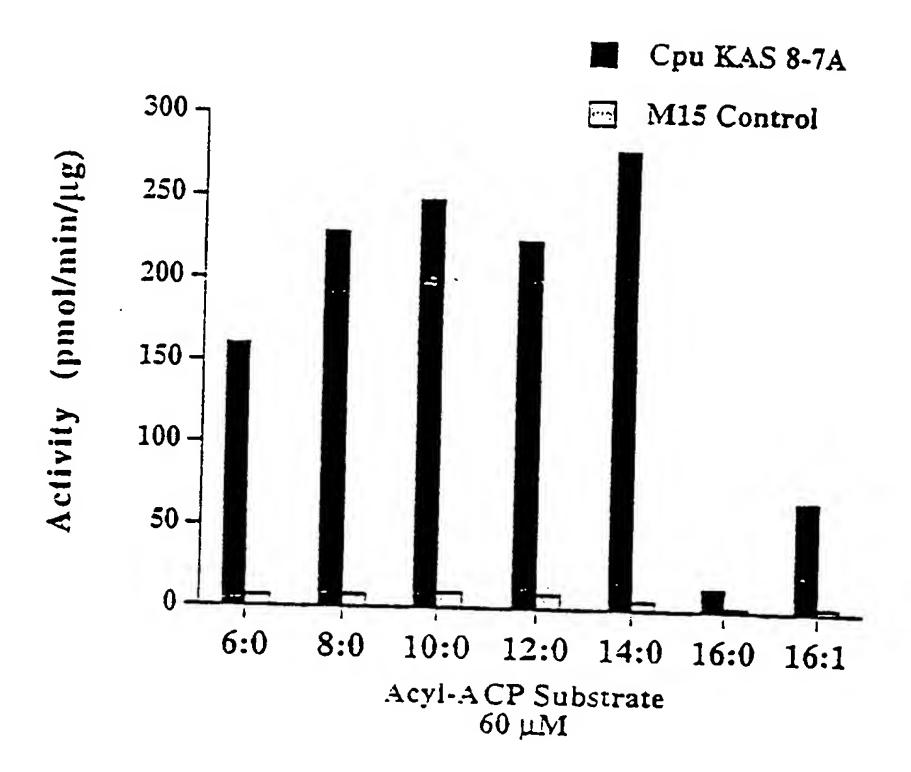


FIGURE 10

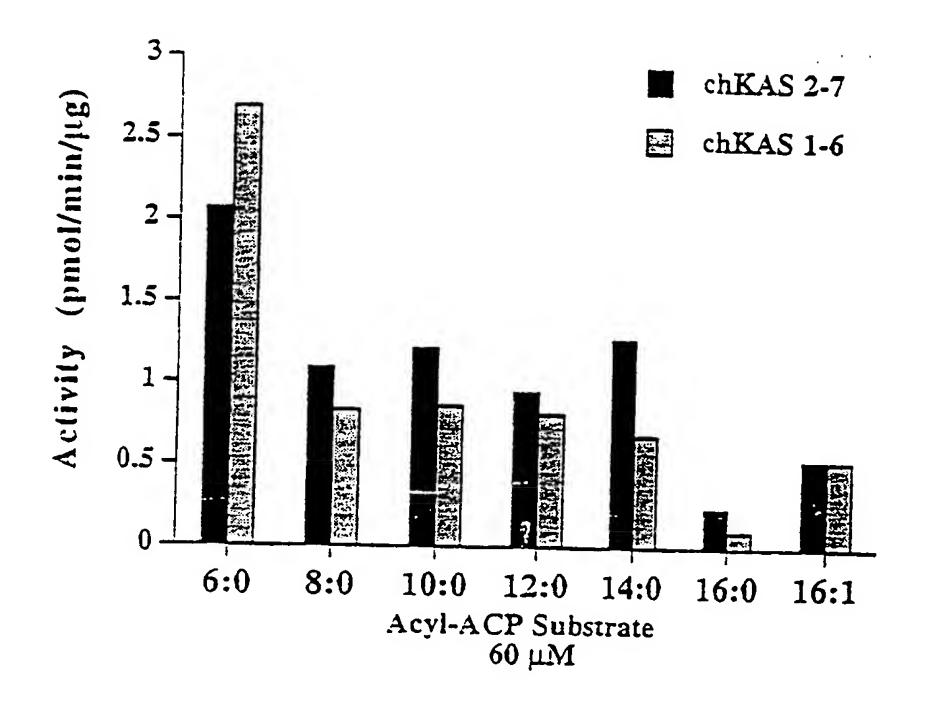


FIGURE 11

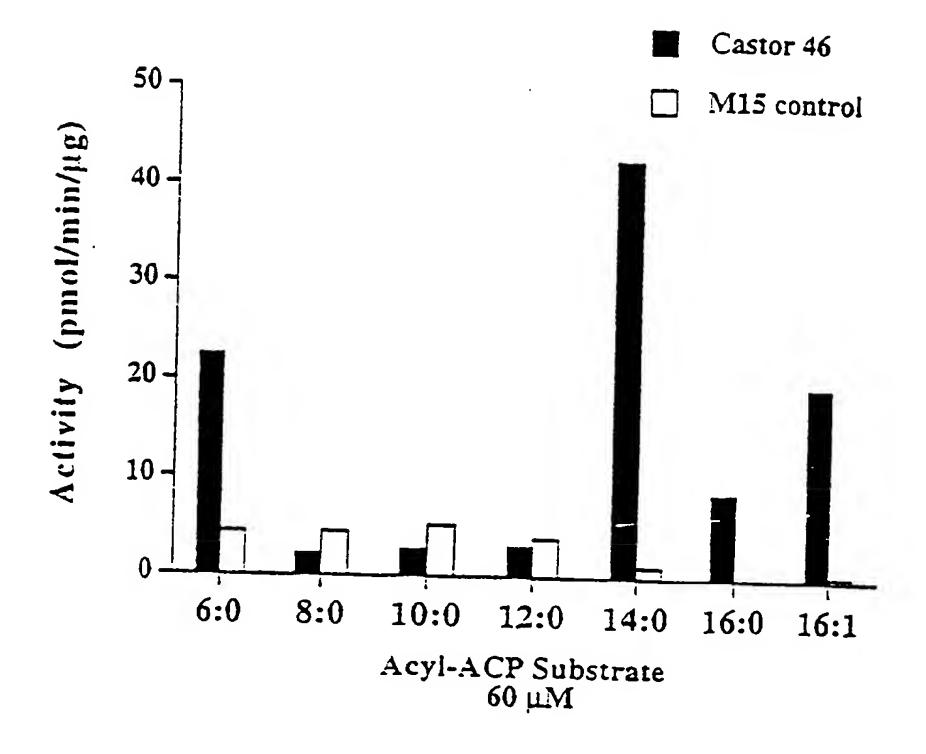
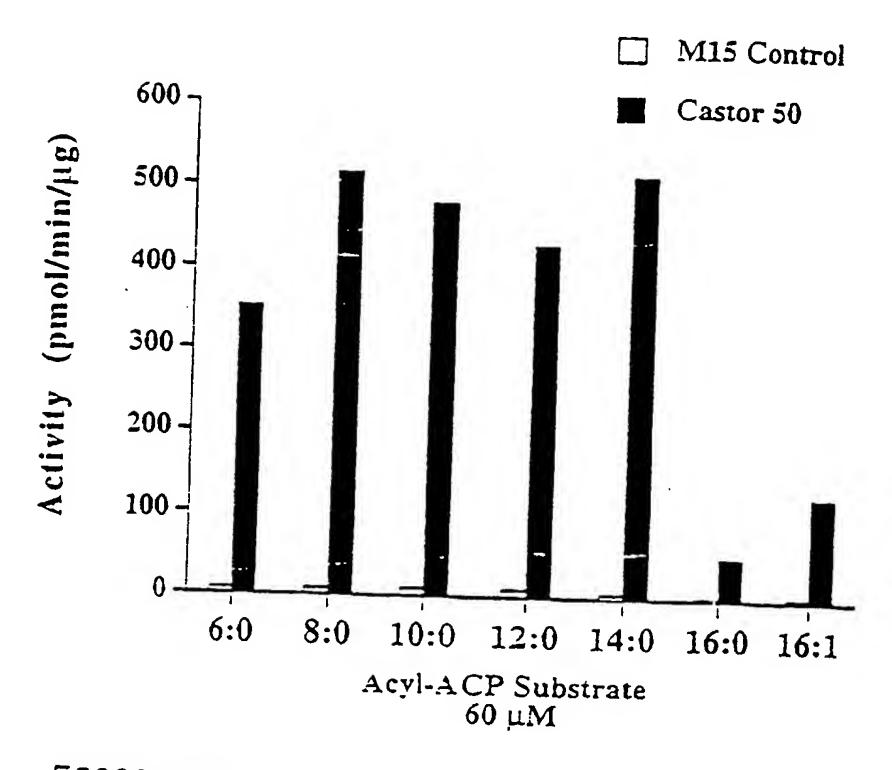


FIGURE 12



E328013-28

FIGURE 13

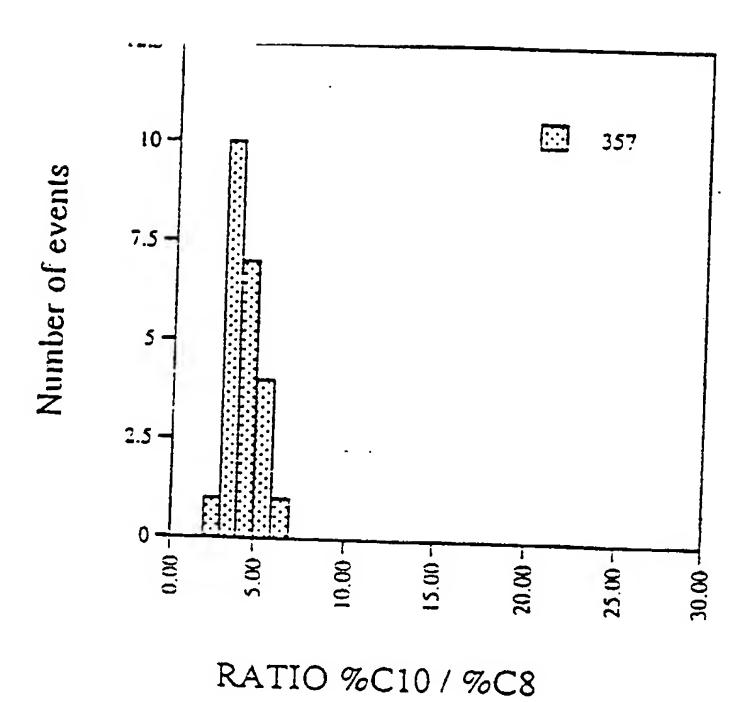


FIGURE 15

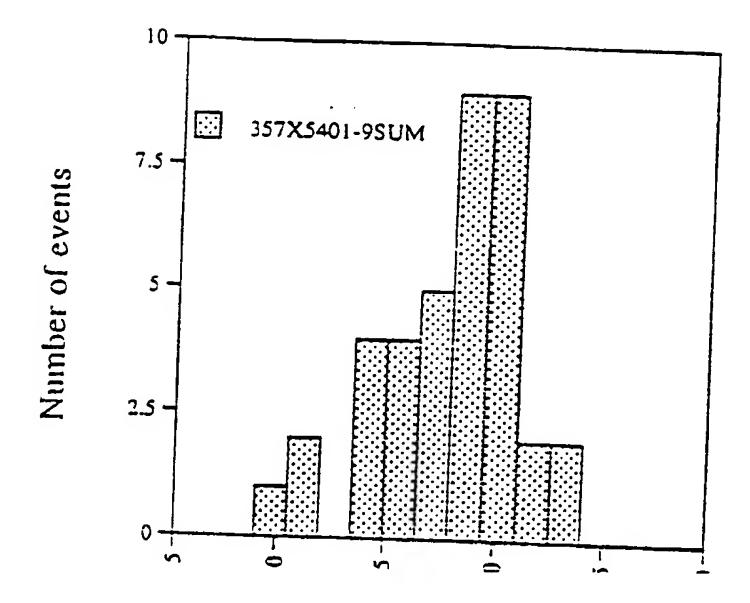
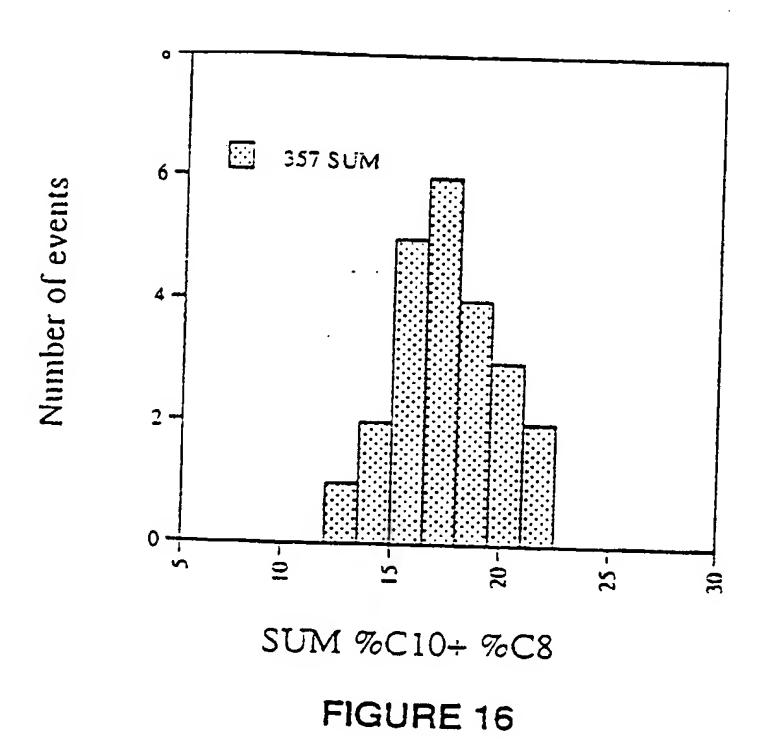


FIGURE 15 2/2



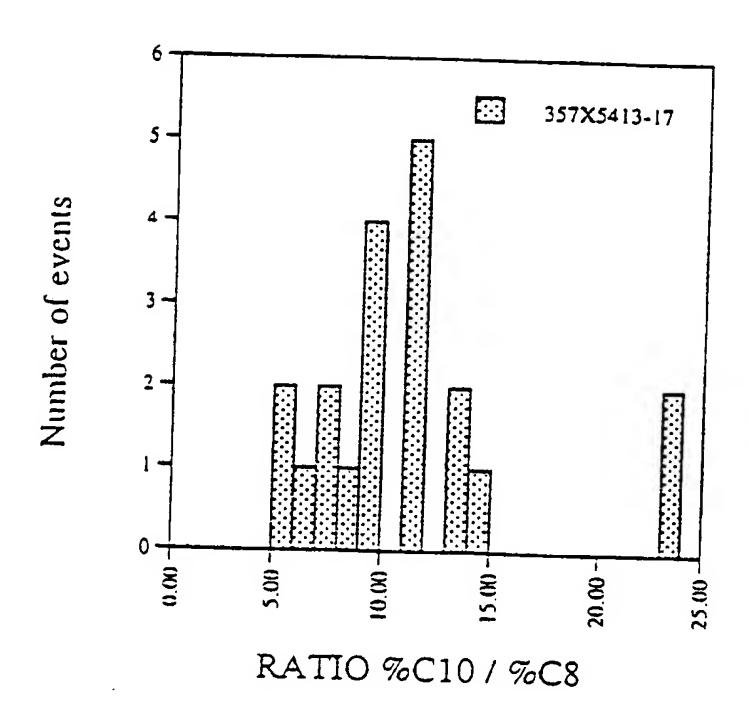


FIGURE 17

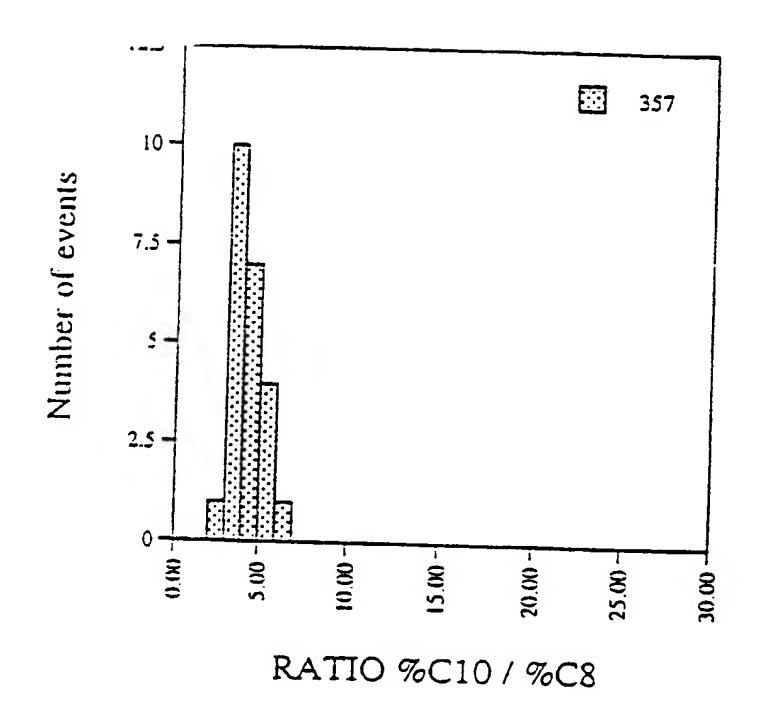
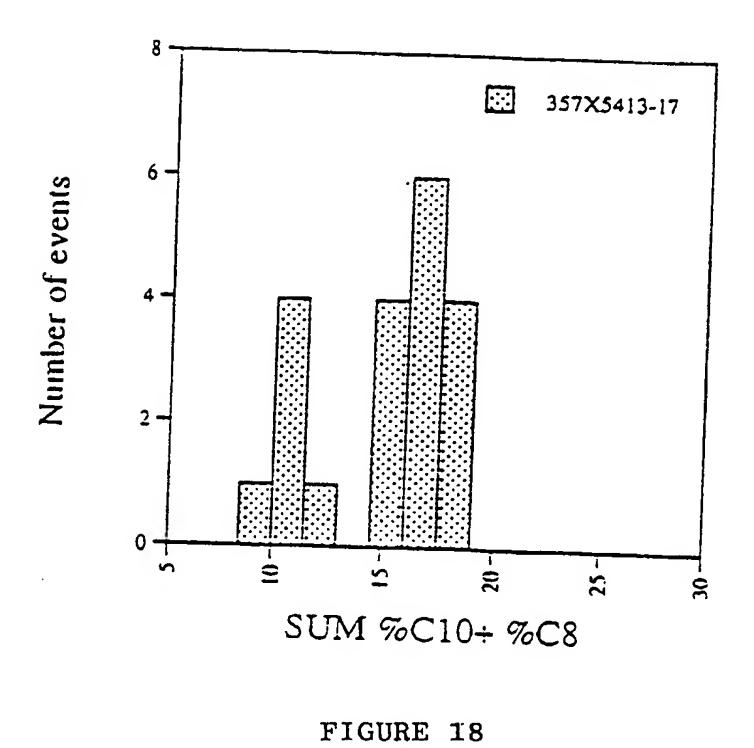
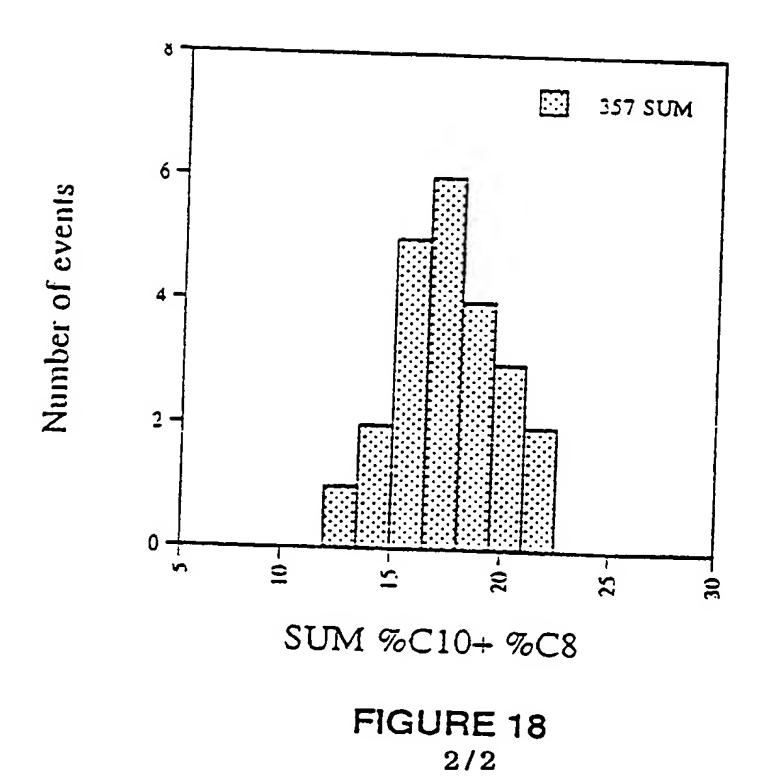


FIGURE 17 2/2



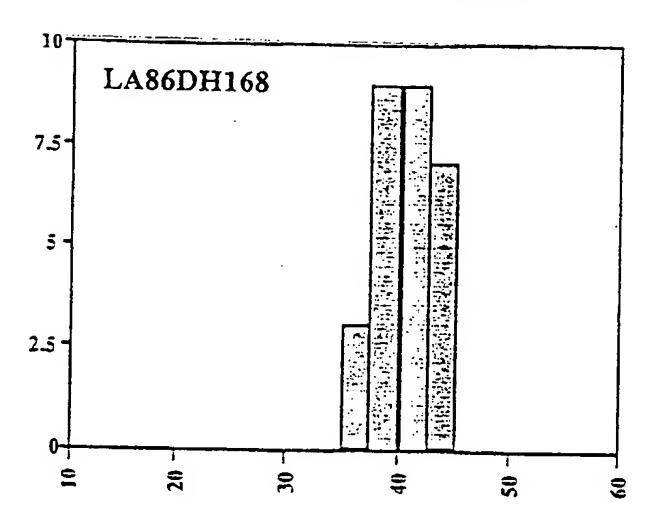
1/2



SUBSTITUTE SHEET (RULE 26)

59/66





12:0 levels (w%)

FIGURE 19 1/3

SUBSTITUTE SHEET (RULE 26)

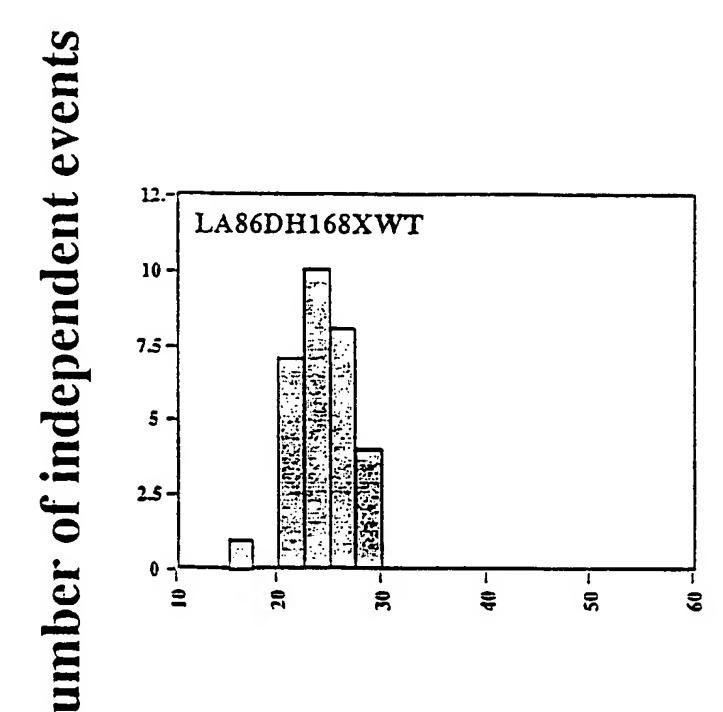
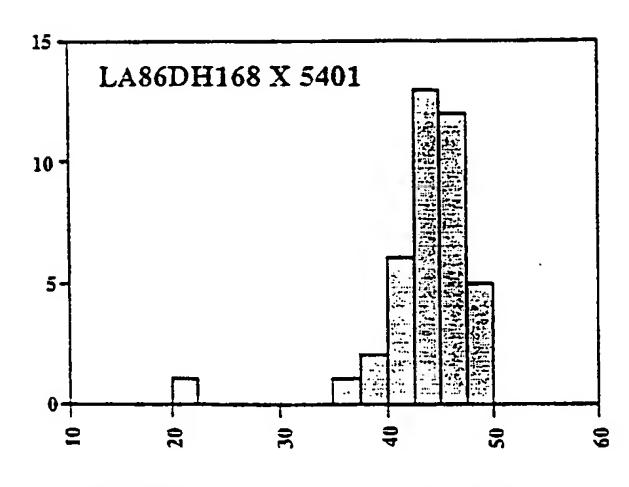


FIGURE 19 3/3 SUBSTITUTE SHEET (RULE 26)

lumber of independent events



12:0 levels (w%)

FIGURE 19 2/3.

SUBSTITUTE SHEET (RULE 26)

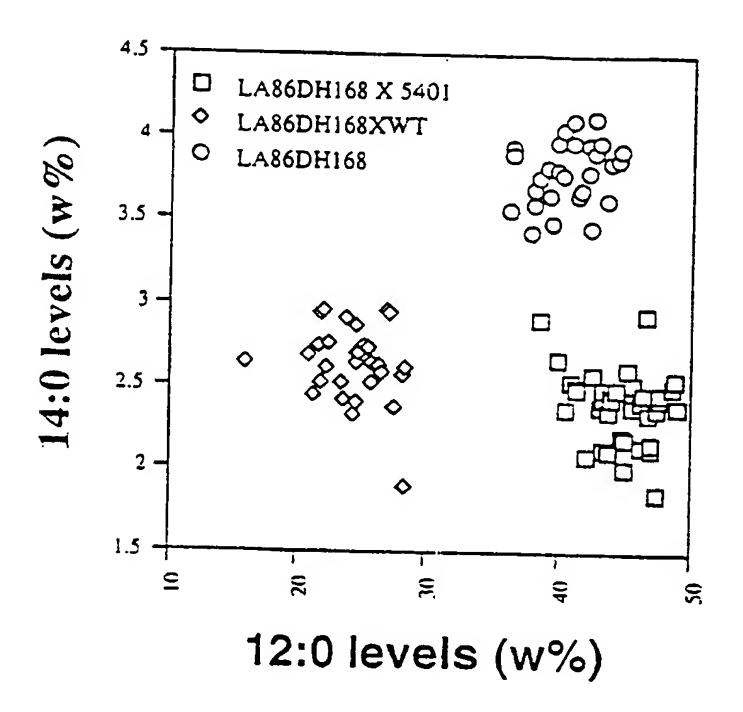
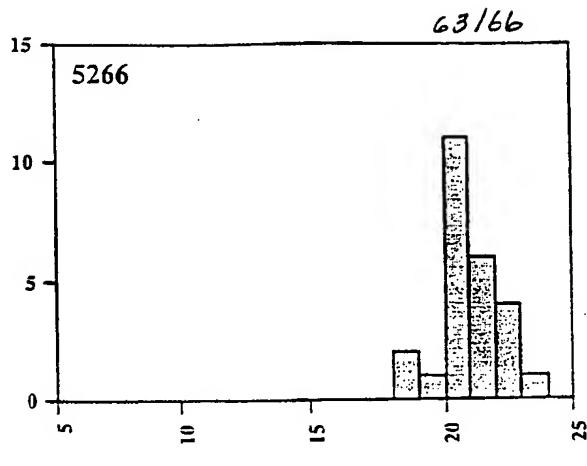


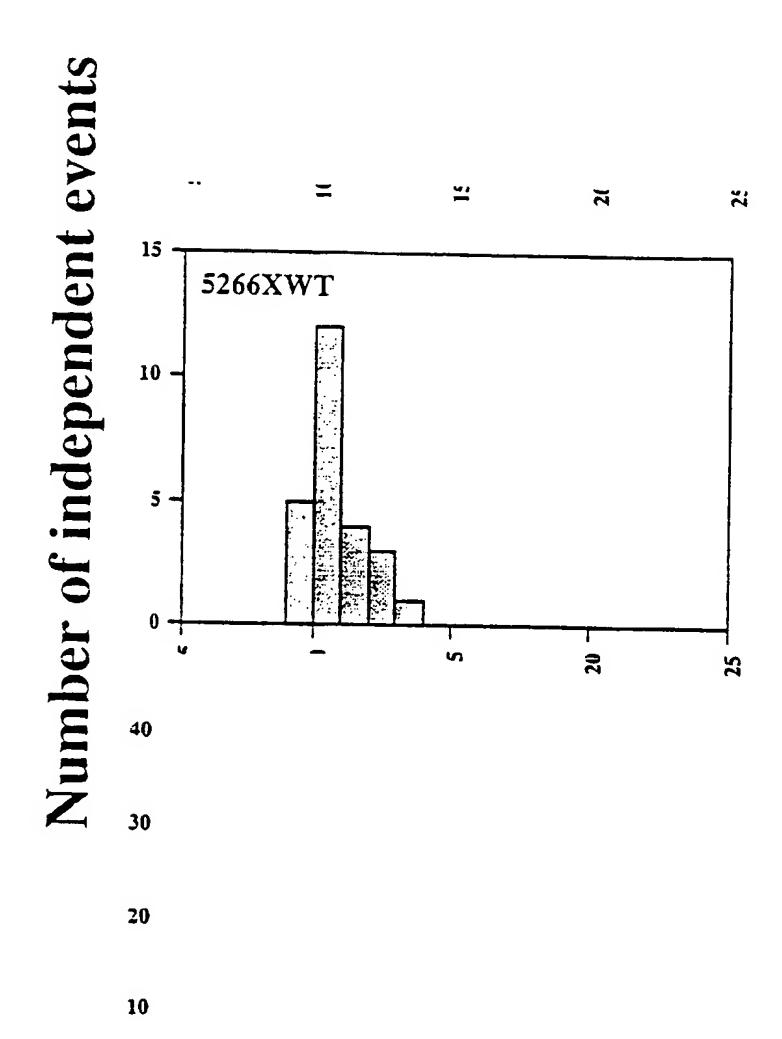
FIGURE 20





18:0 levels (w%)

FIGURE --21. 1/3



18:0 levels (w%)

0

FIGURE 21.

SUBSTITUTE SHEET (RULE 26)

Number of independent events

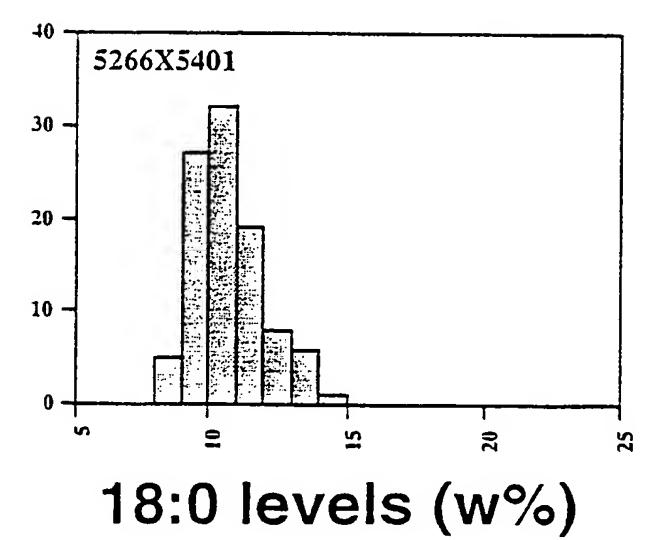


FIGURE 21 3/3

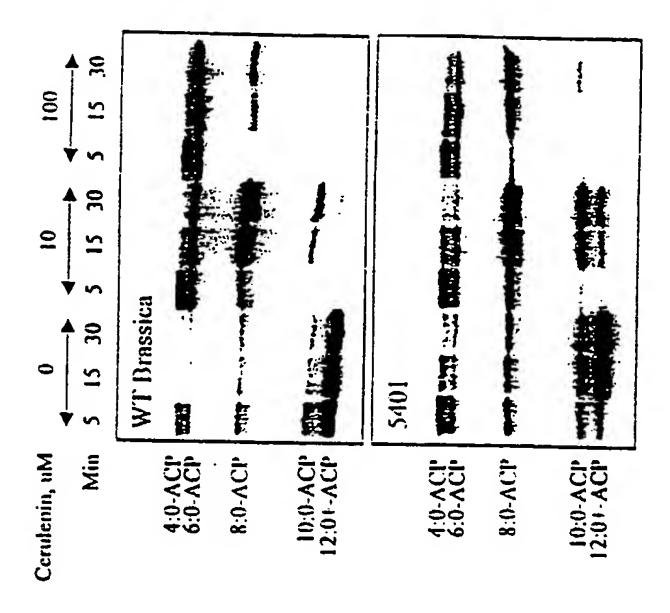


FIGURE 22

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INTERNATIONAL APPLICATION PUBLISI	HED U	INDER THE PATENT COOPERATION TREATY (PCT)
(51) International Patent Classification 6:		(11) International Publication Number: WO 98/46776
C12N 15/82, 15/54	A3	(43) International Publication Date: 22 October 1998 (22.10.98)
(21) International Application Number: PCT/US9 (22) International Filing Date: 9 April 1998 (6)		patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR,
(30) Priority Data: 60/041,815 11 April 1997 (11.04.97)	ι	Published With international search report.
(71) Applicant (for all designated States except US): CALC [US/US]; 1920 Fifth Street, Davis, CA 95616		
(72) Inventor; and (75) Inventor/Applicant (for US only): DEHESH, [US/US]; 521 Crownpointe Circle, Vacaville, C (US).		
(74) Agent: SCHWEDLER, Carl, J.; Calgene LLC, 19 Street, Davis, CA 95616 (US).	920 Fii	th
(54) Title: PLANT FATTY ACID SYNTHASES AND I FATTY ACIDS	USE IN	IMPROVED METHODS FOR PRODUCTION OF MEDIUM-CHAIN
(57) Abstract		
from Cuphea species. Amino acid and nucleic acid for synin constructs for production of genetically engineered plan	nthase plass of plant	to β -ketoacyl-ACP synthase of special interest are synthases obtainable protein factors are provided, as well as methods to utilize such sequences an altered fatty acid compositions. Of particular interest is the expression medium-chain acyl-ACP thioesterases for production of increased levels asgenic plant seeds.

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Inte. ..lonal Application No PCT/US 98/07114

A. CLASSII IPC 6	FICATION OF SUBJECT MATTER C12N15/82 C12N15/54	-	
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X Furt	ther documents are listed in the continuation of box C.	X Patent family members are listed	in annex.
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		PCT/US 98/07114
	ation) DOCUMENTS CONSIDERED TO BE RELEVANT	
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International application No.

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BoxI	Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)						
This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:							
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Remark	The additional search fees were accompanied by the applicant's protest. No protest accompanied the payment of additional search fees.						
	140 protest accompanied the payment of additional search rees.						

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

Claims Nos.: 1-14,19,20,21,26,27,28

Remark: Claims 1-14 were not provided to the ISA at the time of search and hence the subject matter of these claims and the dependent claims 19,20, 21,26,27,28, could not be defined.

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PCT/US 98/07114

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